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The present invention relates to Fc-AGP-1 fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to a genetic or chemical fusion protein comprising the Fc immunoglobulin region, derivative or analog fused to the N-terminal portion of the AGP-1 protein, derivative or analog.

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#### AGP-1 FUSION PROTEIN COMPOSITIONS AND METHODS

## Field of the Invention

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The present invention relates to AGP-1 fusion protein compositions and methods of preparation and use thereof.

# Background of the Invention

The availability of recombinant proteins for therapeutic use has led to advances in protein modifications in order to enhance or improve the properties of such proteins as pharmaceutical agents. Such modifications can provide enhanced protein protection and decreased degradation by reducing or eliminating proteolysis. Additional advantages include, under certain circumstances, increasing the stability, circulation time, and the biological activity of the therapeutic protein. A review article describing protein modifications is Francis, Focus on Growth Factors 3:4-10 (May 1992) (published by Mediscript, London, UK).

One such modification is the use of an Fc region of an immunoglobulin molecule. Antibodies comprise two functionally independent parts, a variable domain known as "Fab", which binds antigen, and a constant domain, known as "Fc" which provides the link to effector functions such as complement or phagocytic cells. The Fc portion of an immunoglobulin has a long plasma half-life, whereas the Fab is short-lived. (Capon, et al., Nature 337, 525-531 (1989)).

Therapeutic protein products have been constructed using the Fc domain to provide longer half-life or to incorporate functions such as Fc receptor binding, protein A binding, complement fixation and placental transfer which all reside in the Fc proteins

of immunogobulins. Id. For example, the Fc region of an IgG1 antibody has been fused to the N-terminal end of CD30 ligand (CD30-L), a molecule which binds CD30 receptors expressed on Hodgkin's Disease tumor cells, anaplastic lymphoma cells, T-cell leukemia cells and other malignant cell types. See, U.S. Patent No. 5,480,981. IL-10, an anti-inflammatory and antirejection agent has been fused to murine Fcy2a in order to increase the cytokines short circulating halflife. (Zheng et al., The Journal of Immunology, 154, 10 5590-5600 (1995)). Studies have also evaluated the use of tumor necrosis factor receptor linked with the Fc protein of human IgG1 to treat patients with septic shock. (Fisher et al., N. Engl. J. Med., 334: 1697-1702 (1996); Van Zee et al., The Journal of Immunology, 15 156: 2221-2230 (1996)). Fc has also been fused with CD4 receptor to produce a therapeutic protein for treatment of AIDS. See, Capon et al., Nature, 337:525-531 (1989). In addition, the N-terminus of interleukin-2(IL-2) has also been fused to the Fc 20 portion of IgG1 or IgG3 to overcome the short half life of IL-2 and its systemic toxicity. See, Harvill et al., Immunotechnology, 1, 95-105 (1995).

necrosis factor (TNF) family has been identified and observed to induce apoptosis in certain tumor cell lines, but not in normal cell lines. This protein has been referred to as TRAIL (Wiley et al. Immunity 3, 673-682 (1995)), Apo-2 (Pitti et al. J. Biol. Chem. 22, 12687-12690 (1996) and AGP-1 (WO97/46686) and is referred to herein as AGP-1. AGP-1 has been studied as a soluble protein which lacks transmembrane and intracellular domains and is most advantageously used as a therapeutic in this form. However, the apoptotic

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activity of soluble AGP-1 is too low for it to be useful as a therapeutic.

Consequently, there exists a need to develop AGP-1 protein compositions for clinical application. Such development would include AGP-1 protein compositions which achieve increased biological activity, decreased degradation, increased stability and increased circulation time. The present invention provides such compositions.

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#### Summary of the Invention

The present invention relates to Fc-AGP-1 fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to a fusion protein 15 comprising an Fc protein, or variant, fragment or derivative thereof, fused to the N-terminal portion of an AGP-1 protein, or variant, fragment, or derivative thereof. Unexpectedly, as described herein, a fusion of an Fc protein to the N-terminus of a soluble AGP-1 20 protein demonstrates enhanced biological activity compared to an unmodified soluble AGP-1 protein. Such unexpected advantages from the Fc modification to AGP-1 protein would be advantageous in that these changes contribute to lower doses required or less frequent dosing. Thus, as described below in more detail, the present invention has a number of aspects relating to the modification of proteins via fusion of an Fc region to an AGP-1 protein (or variant, fragments or derivative thereof), as well as, specific 30 modifications, preparations and methods of use thereof.

The present invention provides for a protein having a formula selected from the group consisting of:  $R_1 - R_2$  and  $R_1 - L - R_2$ , wherein  $R_1$  is a Fc protein, or a variant or fragment thereof,  $R_2$  is an AGP-1 protein, or variant or fragment thereof, and L is a linker. The

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invention provides for genetic or chemical linkages of the R1 and R2 moieties as described herein.

In one aspect, the present invention provides a Fc-AGP-1 fusion protein wherein Fc (or a variant, fragment or derivative thereof) is genetically fused to the N-terminus of an AGP-1 protein (or a variant, fragment or derivative thereof). In another aspect of the invention, an Fc portion may also be linked to the N-terminus of an AGP-1 protein (or a variant, fragment or derivative thereof) by a peptide or chemical linker as known in the art. As noted above and described in more detail below, the Fc-AGP-1 fusion protein has unexpected enhanced biological activity when compared to a soluble AGP-1 protein. Additional aspects of the present invention, therefore, include not only Fc-AGP-1 fusion protein compositions, but also nucleic acid sequences encoding such proteins, related vectors and host cells containing such vectors, both useful for producing fusion proteins of the present invention.

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In a second aspect, the present invention provides for preparing the Fc-AGP-1 fusion protein. Such methods include recombinant DNA techniques for preparation of recombinant proteins. Furthermore, such aspects include methods of protein production and purification as well.

In another aspect, the present invention provides methods for treating proliferative disorders, such as cancer or cardiovascular diseases, viral infections and viral-induced diseases, and immune disorders by administration of Fc-AGP-1 fusion proteins.

In another aspect, the present invention also provides for related pharmaceutical compositions of the Fc-AGP-1 proteins, variants, fragments and derivatives thereof, for use in the above therapies.

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# Description of the Figures

Figure 1 (SEQ ID NO: 32) shows the amino acid sequence of the hinge, CH2 and CH3 regions of human  $IgG\gamma 1$ .

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Figure 2 (SEQ ID NOS: 33 and 34) shows the nucleotide and amino acid sequence of human AGP-1.

Figure 3 (SEQ ID NOS: 35 and 36) shows the nucleotide and amino acid sequence of Fc-huAGP-1 (95-281). Amino acids corresponding to OPG signal peptide are underlined. Amino acids corresponding to human AGP-1 (95-281) are bracketed.

15 Figure 4 (SEQ ID NOS: 37 and 38) shows the nucleotide and amino acid sequence of Fc-huAGP-1 (114-281). Amino acids corresponding to OPG signal peptide are underlined. Amino acids corresponding to human AGP-1 (114-281) are bracketed.

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Figure 5 (SEQ ID NOS: 39 and 40) shows the nucleotide and amino acid sequence of Fc-muAGP-1 (99-291). Amino acids corresponding to OPG signal peptide are underlined. Amino acids corresponding to murine AGP-1 (99-291) are bracketed.

Figure 6 (SEQ ID NOS: 41 and 42) shows the nucleotide and amino acid sequence of Fc-muAGP-1 (120-291). Amino acids corresponding to OPG signal peptide are underlined. Amino acids corresponding to murine AGP-1(120-291) are bracketed.

Figure 7 shows the activity of soluble AGP-1 and Fc-AGP-1 fusion protein in inducing apoptosis in cultured Jurkat cells.

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# Detailed Description of the Invention

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The present invention relates to Fc-AGP-1 fusion protein compositions, methods of preparation of such compositions and uses thereof. In particular, the present invention relates to the genetic or chemical fusion of the Fc region of immunoglobulins to the N-terminal portion of the AGP-1 protein. Unexpectedly, fusion of Fc at the N-terminus of the AGP-1 protein demonstrates advantages which are not seen in soluble AGP-1 protein. Surprisingly, the N-terminally modified Fc-AGP-1 protein provides unexpected increased biological activity. Accordingly, the Fc-AGP-1 fusion protein, and variants, fragments and derivatives thereof, as well as, related methods of use and preparation, are described in more detail below.

The term "Fc" refers to a molecule or sequence comprising the sequence of a non-antigenbinding portion of antibody, whether in monomeric or multimeric form. The original immunoglobulin source of an Fc is preferably of human origin and may be from any isotype, e.g., IgG, IgA, IgM, IgE or IgD. One method of preparation of an isolated Fc molecule involves digestion of an antibody with papain to separate antigen and non-antigen binding portions of the antibody. Another method of preparation of an isolated Fc molecules is production by recombinant DNA expression followed by purification of the Fc molecules so expressed. A full-length Fc consists of the following Ig heavy chain regions: CH1, CH2 and CH3 wherein the CH1 and CH2 regions are typically connected by a flexible hinge region. In one embodiment, an Fc has the amino acid sequence of IgG1 such as that shown in Figure 1. The terms "Fc protein, "Fc sequence", "Fc molecules, "Fc region" and "Fc portion" are taken to 35 have the same meaning as "Fc".

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The term "fragment" when used in association with Fc or AGP-1 polypeptides, or fusion polypeptides thereof, refers to a peptide or polypeptide that comprises less than the full length amino acid sequence of an Fc or AGP-1 polypeptide. Such a fragment may arise, for example, from a truncation at the amino terminus, a truncation at the carboxy terminus, and/or an internal deletion of a residue(s) from the amino acid sequence. AGP-1 or Fc fragments may result from alternative RNA splicing or from in vivo protease activity.

The term "variant" when used in association with Fc or AGP-1 polypeptides, or with fusion polypeptides thereof, refers to a polypeptide comprising an amino acid sequence which contain one or more amino acid sequence substitutions, deletions, and/or additions as compared to native Fc or AGP-1 polypeptide amino acid sequences. Variants may be naturally occurring or artificially constructed. Variants of the invention may be prepared from the corresponding nucleic acid molecules encoding said variants, which have a DNA sequence that varies accordingly from the DNA sequences for native Fc or AGP-1 polypeptides.

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The term "derivative" when used in association with Fc or AGP-1 polypeptides, or with fusion polypeptides thereof, refers to Fc or AGP-1 polypeptide variants or fragments thereof, that have been chemically modified, as for example, by covalent attachment of one or more polymers, including, but limited to, water soluble polymers, N-linked or O-linked carbohydrates, sugars, phosphates, and/or other such molecules. The derivatives are modified in a manner that is different from native Fc or AGP-1, either in the type or location of the molecules attached to the polypeptide. Derivatives further

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includes deletion of one or more chemical groups naturally attached to an Fc or AGP-1 polypeptide.

The term "fusion" refers to joining of different peptide or protein segments by genetic or chemical methods wherein the joined ends of the peptide or protein segments may be directly adjacent to each other or may be separated by linker or spacer moieties such as amino acid residues or other linking groups.

#### 10 Compositions

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The invention provides for FcAGP-1 fusion polypeptides and compositions thereof. Fusions of an Fc region to an AGP-1 polypeptide are advantageously made at the amino terminus of AGP-1, that is, the carboxy terminus of an Fc region is fused to the amino terminus of AGP-1. These fusion proteins (and nucleic acids encoding same) are designated herein as FcAGP-1. However, it is also contemplated that, in certain instances, it may be desirable to fuse the carboxy terminus of AGP-1 to the amino terminus of an Fc region. The fusion proteins (and nucleic acids encoding same) are designated herein as AGP-1Fc.

An Fc, or a variant, fragment or derivative thereof, may be from an Ig class. In one embodiment, an Fc is from the IgG class, such as IgG1, IgG2, IgG3, and IgG4. In another embodiment, an Fc is from IgG1. An Fc may also comprise amino acid residues represented by a combination of any two or more of the Ig classes, such as residues from IgG1 and IgG2, or from IgG1, IgG2 and IgG3, and so forth. In one embodiment, an Fc region of an Fc-AGP-1 fusion protein has the sequence as set forth in Figure 1 (SEQ ID NO: 32) comprising hinge CH2 and CH3 regions of human IgG1. (see Ellison et al., Nucleic Acids Res. 10, 4071-4079 (1982).

In addition to naturally occurring variations in Fc regions, Fc variants, fragments and derivatives

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may contain non-naturally occurring changes in Fc which are constructed by, for example, introducing substitutions, additions, insertions or deletions of residues or sequences in a native or naturally occurring Fc, or by modifying the Fc portion by chemical modification and the like. In general, Fc variants, fragments and derivatives are prepared such that the increased circulating half-life of Fc fusions to AGP-1 is largely retained.

Also provided by the invention are Fc variants with conservative amino acid substitutions. The term "conservative amino acid substitution" refers to a substitution of a native amino acid residue with a nonnative residue such that there is little or no effect on the polarity or charge of the amino acid residue at that position. For example, a conservative substitution results from the replacement of a non-polar residue in a polypeptide with any other non-polar residue. General rules for conservative amino acid substitutions are set forth in Table I.

Table I
Conservative Amino Acid Substitutions

Original Residues	Exemplary Substitutions	Preferred Substitutions
Ala	Val,Leu,Ile	Val
Arg	Lys,Gln,Asn	Lys
Asn	Gln, His, Lys, Arg	Gln
Asp	Glu	Glu
Cys	Ser	Ser
Gln	Asn	Asn
Glu	Asp	Asp
Gly	Pro,Ala	Ala
His	Asn, Gln, Lys, Arg	Arg
Ile	Leu, Val, Met, Ala,	Leu
	Phe, Norleucine	

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Leu	Norleucine, Ile,	Ile
	Val, Met, Ala, Phe	
Lys	Arg, Gln, Asn	Arg
Met	Leu, Phe, Ile	Leu
Phe	Leu, Val, Ile, Ala,	Leu
	Tyr	
Pro	Ala	Ala
Ser	Thr	Thr
Thr	Ser	Ser
Trp	Tyr, Phe	Tyr
Tyr	Trp, Phe, Thr, Ser	Phe
Val	Ile, Met, Leu, Phe,	Leu
	Ala, Norleucine	

Conservative amino acid substitutions also encompass non-naturally occurring amino acid residues which are typically incorporated by chemical peptide synthesis rather than by synthesis in biological systems. These include peptidomimetics, and other reversed or inverted forms of amino acid moieties. Conservative modifications to the amino acid sequence (and the corresponding modifications to the encoding nucleotides) are expected to produce Fc molecules (and 10 FcAGP-1 fusion proteins) having functional and chemical characteristics similar to those of unmodified Fc and

In addition to the substitutions set forth in Table I, any native residue in an Fc region (or in an 15 FcAGP-1 fusion protein) may also be substituted with alanine, as has been previously described for "alanine scanning mutagenesis" (Cunningham et al. Science 244, 1081-1085 (1989)).

FcAGP-1 proteins.

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Substantial modifications in the functional and/or chemical characteristics of an Fc molecule (and an FcAGP-1 fusion protein) may be accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the 25 molecular backbone in the area of the substitution, for

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example, as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side chain. Naturally occurring residues may be divided into groups based on common side chain properties:

- hydrophobic: norleucine, Met, Ala, Val, Leu,
   Ile;
- 2) neutral hydrophilic: Cys, Ser, Thr;
- 3) acidic: Asp, Glu;
- 10 4) basic: Asn, Gln, His, Lys, Arg;
  - 5) residues that influence chain orientation: Gly, Pro; and
  - 6) aromatic: Trp, Tyr, Phe.

Non-conservative substitutions may involve

the exchange of a member of one of these classes for a
member from another class. Such substituted residues
may be introduced into regions of an Fc or AGP-1
molecule that are homologous with non-human Fc or AGP1, or into the non-homologous regions of the molecule.

20 Cysteine residues in Fc molecules can be deleted or replaced with other amino acids to prevent formation of disulfide crosslinks. In particular, a cysteine residue at position 5 of Figure 1 (SEQ. ID. NO. 32) may be substituted with one or more amino acids, such as alanine or serine. Alternatively, the cysteine residue at position 5 could be deleted.

An Fc fragment may be prepared by deletion of one or more amino acids at any of positions 1, 2, 3, 4 and 5 as shown in Figure 1 (SEQ ID NO. 32). In one embodiment, the amino acid residues at positions 1-5 inclusive are deleted. Substitutions at these positions can also be made and are with in the scope of this invention.

Fc variants may also be made which show

reduced binding to Fc receptors which trigger effector functions such as antibody dependent cellular

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cytotoxicity (ADCC) and activation of complement. Such variants may include leucine at position 20 deleted or substituted with a glutamine residue, glutamate at position 103 deleted or substituted with an alanine residue, and lysines at positions 105 and 107 deleted or substituted with alanine residues (following the numbering as set forth in Figure 1). One or more of such substitutions are contemplated.

In one embodiment, Fc variants will exhibit stronger binding to the FcRn receptor ("salvage 10 receptor") and a longer circulating half-life compared to native Fc such as that shown in Figure 1. Example of such variants include amino acid substitutions at one or more of residues 33, 35-42, 59, 72, 75, 77, 95-98, 101, 172-174, 215 and 220-223, wherein the substitution(s) confer tighter binding of an Fc variant to the FcRn receptor.

Other Fc variants include one or more tyrosine residues replaced with, for example, phenyalanine residues. In addition, other variant amino acid insertions, deletions and/or substitutions are also contemplated and are within the scope of the present invention. Examples include Fc variants disclosed in WO96/32478 and WO97/34630 hereby incorporated by reference. Furthermore, alterations may be in the form of altered amino acids, such as peptidomimetics or D-amino acids.

The Fc protein may be also linked to the AGP-1 proteins of the Fc-AGP-1 protein by "linker" moieties whether chemical or amino acids of varying lengths. Such chemical linkers are well known in the art. Amino acid linker sequences can include but are not limited to:

- ala-ala-ala; (a)
- ala-ala-ala; 35 (b)

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ala-ala-ala-ala; (c)

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	(d)	gly-gly;
	(e)	gly-gly-gly;
	(f)	gly-gly-gly-gly;
	(g)	gly-gly-gly-gly-gly-gly;
5	(h)	gly-pro-gly;
	(i)	gly-gly-pro-gly-gly; and
	(j)	any combination of subparts (a)
	through (i).	

AGP-1 variants, fragments and derivatives are 10 also provided by the invention and are generally as described hereinabove for Fc molecules, with the exception of the specific locations of the modified amino acid residues. In a preferred embodiment, AGP-1 is a soluble form of AGP-1 which is not membrane-bound 15 and lacks a functional transmembrane domain. As an example, soluble AGP-1 may comprise an extracellular domain and lack sequences for cytoplasmic and transmembrane domains. The full-length human AGP-1 extracellular domain encompasses about residues 39-281 20 inclusive using the numbering system as set forth in Figure 2. Soluble human AGP-1 may also encompass fragments of a full-length extracellular domain which function to bind receptor or to induce apoptosis in an assay such as that described in Example 4 below. 25 Soluble AGP-1 fragments comprise the amino acid sequence X-281 wherein X is any residue from 95 to 114 inclusive using the numbering system of as set forth in Figure 2. Other soluble AGP-1 fragments encompass residues 115-281, 116-281, 117-281, 118-281 and 119-281 30 inclusive. Variants and derivatives of the AGP-1 fragments described herein are also encompassed by the invention.

# 35 Nucleic acid molecules

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Nucleic acid molecules encoding Fc-AGP-1 proteins, or variants, fragments or derivatives thereof, are provided for by the invention. Nucleic acid molecules of the invention may be produced using site directed mutagenesis, PCR amplification, or other 5 appropriate methods, where the primer(s) have the desired mutations. See Sambrook et al. (Molecular Cloning: A Laboratory Manual, Cold Springs Harbor Laboratory Press, Cold Springs Harbor, N.Y. (1989)), and Ausubel et al. (Current Protocols in Molecular 10 Biology, Wiley and Sons, N.Y. (1994)), for descriptions of mutagenesis techniques. Chemical synthesis using methods described by Engels et al. (Angew. Chem. Intl. Ed. 28, 716-734 (1989)), may also be used to prepare such variants. Other methods known to the skilled 15 artisan may be used as well.

In certain embodiments, nucleic acid variants contain codons which have been altered for optimal expression of an Fc-AGP-1 polypeptide in a given host cell. Particular codon alterations will depend upon the Fc-AGP-1 polypeptide(s) and host cell(s) selected for expression. Such "codon optimization" can be carried out by a variety of methods, for example, by selecting codons which are preferred for use in highly expressed genes in a given host cell. Computer algorithms which incorporate codon frequency tables such as "Ecohigh. Cod" for codon preference of highly expressed bacterial genes may be used and are provided by the University of Wisconsin Package Version 9.0, Genetics Computer Group, Madison, WI. Other useful codon frequency tables include "Celegans\_high.cod", "Celegans\_low.cod", "Drosophila\_high.cod", "Human\_high.cod", "Maize\_high.cod", and "Yeast\_high.cod".

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In other embodiments, nucleic acid molecules encode Fc-AGP-1 variants with conservative amino acid

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substitutions as defined hereinabove. Also provided for are Fc or AGP-1 variants comprising an addition and/or a deletion of one or more N-linked or O-linked glycosylation sites, or Fc or AGP-1 polypeptide fragments as described above. In addition, nucleic acid molecules may encode any combination of Fc and/or AGP-1 variants, fragments, and fusion polypeptides described herein.

#### 10 Vectors and Host cells

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A nucleic acid molecule encoding an Fc-AGP-1 fusion protein is inserted into an appropriate expression vector using standard ligation techniques. The vector is typically selected to be functional in the particular host cell employed (i.e., the vector is 15 compatible with the host cell machinery such that amplification of the gene and/or expression of the gene can occur). A nucleic acid molecule encoding an Fc-AGP-1 protein may be amplified/expressed in prokaryotic, yeast, insect (baculovirus systems) and/or 20 eukaryotic host cells. Selection of the host cell will depend in part on whether an Fc-AGP-1 protein is to be post-translationally modified (e.g, glycosylated and/or phosphorylated). If so, yeast, insect, or mammalian 25 host cells are preferable.

Typically, expression vectors used in any of the host cells will contain sequences for plasmid maintenance and for cloning and expression of exogenous nucleotide sequences. such sequences, collectively referred to as "flanking sequences" in certain embodiments will typically include one or more of the following nucleotides: a promoter, one or more enhancer sequences, an origin of replication, a transcriptional termination sequence, a complete intron sequence containing a donor and acceptor splice site, a leader sequence for secretion, a ribosome binding site,

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a polyadenylation sequence, a polylinker region for inserting the nucleic acid encoding the polypeptide to be expressed, and a selectable marker element.

Flanking sequences may be homologous (i.e.,

from the same species and/or strain as the host cell),
heterologous (i.e., from a species other than the host
cell species or strain), hybrid (i.e., a combination of
flanking sequences from more than one source), or
synthetic, or native sequences which normally function
to regulate AGP-1 and/or Fc protein expression. As
such, the source of flanking sequences may be any
prokaryotic or eukaryotic organism, any vertebrate or
invertebrate organism, or any plant, provided that the
flanking sequences is functional in, and can be
activated by, the host cell machinery.

A leader, or signal, sequence may be used to direct an Fc-AGP-1 polypeptide out of the host cell. Typically, the signal sequence is positioned in the coding region of the Fc-AGP-1 nucleic acid molecule, or directly at the 5' end of the Fc-AGP-1 polypeptide coding region. Many signal sequences have been identified, and any of them that are functional in the selected host cell may be used in conjunction with nucleic acid sequences encoding Fc-AGP-1 proteins. Therefore, a signal sequence may be homologous

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(naturally occurring) or heterologous to the AGP-1 or Fc gene or cDNA, Additionally, a signal sequence may be chemically synthesized using methods set forth above. In most cases, secretion of an Fc-AGP-1 polypeptide from the host cell via the presence of a signal peptide will result in the removal of the signal peptide from the fusion polypeptide.

The signal sequence may be a component of the vector, or it may be a part of Fc-AGP-1 DNA that is inserted into the vector. Native AGP-1 DNA encodes a signal sequence at the amino terminus of the protein

that is cleaved during post-translational processing of the molecule to form the mature protein (see Figure 2). Included within the scope of this invention are AGP-1 nucleotides with the native signal sequence as well as AGP-1 nucleotides wherein the native signal sequence is deleted and replaced with a heterologous signal sequence. A heterologous signal sequence selected should be one that is recognized and processed, i.e., cleaved by a signal peptidase, by the host cell. In one embodiment, a heterologous signal sequence is the 10 OPG signal sequence as described in WO97/23614. For prokaryotic host cells that do not recognize and process the native AGP-1 signal sequence, the signal sequence is substituted by a prokaryotic signal sequence selected, for example, from the group of the 15 alkaline phosphatase, penicillinase, or heat-stable enterotoxin II leaders. For yeast secretion, the native AGP-1 signal sequence may be substituted by the yeast invertase, alpha factor, or acid phosphatase leaders. In mammalian cell expression the native 20 signal sequence is satisfactory, although other mammalian signal sequences may be suitable.

Preferred vectors for practicing this invention are those which are compatible with

25 bacterial, insect, and mammalian host cells. Such vectors include, inter alia, pCRII, pCR3, and pcDNA3.1 (Invitrogen Company, San Diego, CA), pBSII (Stratagene Company, La Jolla, CA), pET15b (Novagen, Madison, WI), pGEX (Pharmacia Biotech, Piscataway, NJ), pEGFP-N2

30 (Clontech, Palo Alto, CA), pETL (BlueBacII; Invitrogen), pDSR-alpha (PCT Publication No. W090/14363) and pFastBacDual (Gibco/BRL, Grand Island, NY).

Additional possible vectors include, but are not limited to, cosmids, plasmids or modified viruses, but the vector system must be compatible with the

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selected host cell. Such vectors include, but are not limited to plasmids such as Bluescript plasmid derivatives (a high copy number ColE1-based phagemid, Stratagene Cloning Systems Inc., La Jolla CA), PCR cloning plasmids designed for cloning Taq-amplified PCR products (e.g., TOPO™ TA Cloning® Kit, PCR2.1® plasmid derivatives, Invitrogen, Carlsbad, CA), and mammalian, yeast or virus vectors such as a baculovirus expression system (pBacPAK plasmid derivatives, Clontech, Palo Alto, CA). The recombinant molecules can be introduced into host cells via transformation, transfection, infection, electroporation, or other known techniques. After the vector has been constructed and a nucleic acid molecule encoding an AGP-1 polypeptide has been inserted into the proper site of the vector, the completed vector may be inserted into a suitable host cell for amplification and/or polypeptide expression.

(such as *E. coli*) or eukaryotic host cells (such as a yeast cell, an insect cell, or a vertebrate cell). The host cell, when cultured under appropriate conditions, synthesizes an AGP-1 polypeptide which can subsequently be collected from the culture medium (if the host cell secretes it into the medium) or directly from the host cell producing it (if it is not secreted). Selection of an appropriate host cell will depend upon various factors, such as desired expression levels, polypeptide modifications that are desirable or necessary for activity, such as glycosylation or phosphorylation, and ease of folding into a biologically active molecule.

Suitable host cells or cell lines may be mammalian cells, such as Chinese hamster ovary cells (CHO) (ATCC #CCL61 and Urlaub et al., Proc. Natl. Acad. Sci. USA 77, 4216-4220 (1980)), human embryonic kidney (HEK) 293 or 293T cells (ATCC #CRL1573), or 3T3 cells

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(ATCC #CRL1658). The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening and product production and purification are known in the art. Other suitable 5 mammalian cell lines, are the monkey COS-1 and COS-7 cell lines (ATCC #CRL1651), and the CV-1 cell line (ATCC #CCL70). Further exemplary mammalian host cells include primate cell lines and rodent cell lines, including transformed cell lines. Normal diploid cells, cell strains derived from in vitro culture of primary tissue, as well as primary explants, are also suitable. Candidate cells may be genotypically deficient in the selection gene, or may contain a dominantly acting selection gene. Other suitable mammalian cell lines include but are not limited to, mouse neuroblastoma N2A cells, HeLa, mouse L-929 cells, 3T3 lines derived from Swiss, Balb-c or NIH mice, BHK or HaK hamster cell lines. Each of these cell lines is known by and available to those skilled in the art.

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Similarly useful as host cells suitable for the present invention are bacterial cells. For example, the various strains of E. coli (e.g., HB101, DH5a, DH10, and MC1061) are well-known as host cells in the field of biotechnology. Various strains of B. subtilis, Pseudomonas spp., other Bacillus spp., Streptomyces spp., and the like may also be employed in this method.

Many strains of yeast cells known to those skilled in the art are also available as host cells for expression of the polypeptides of the present invention. Preferred yeast cells include, for example, Saccharomyces cerivisae.

Additionally, where desired, insect cell systems may be utilized in the methods of the present invention. Such systems are described for example in Kitts et al. (Biotechniques, 14, 810-817 (1993)),

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Lucklow (Curr. Opin. Biotechnol., 4, 564-572 (1993)) and Lucklow et al. (J. Virol., 67, 4566-4579 (1993)). Preferred insect cells are Sf-9 and Hi5 (Invitrogen, Carlsbad, CA).

Transformation or transfection of an expression vector for an AGP-1 polypeptide into a selected host cell may be accomplished by well known methods including methods such as calcium chloride, electroporation, microinjection, lipofection or the DEAE-dextran method. The method selected will in part be a function of the type of host cell to be used. These methods and other suitable methods are well known to the skilled artisan, and are set forth, for example, in Sambrook et al., supra.

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# Polypeptide Production

Host cells comprising an AGP-1 expression vector (i.e., transformed or transfected) may be cultured using standard media well known to the skilled artisan. The media will usually contain all nutrients necessary for the growth and survival of the cells. Suitable media for culturing E. coli cells are for example, Luria Broth (LB) and/or Terrific Broth (TB). Suitable media for culturing eukaryotic cells are RPMI 1640, MEM, DMEM, all of which may be supplemented with serum and/or growth factors as required by the particular cell line being cultured. A suitable medium for insect cultures is Grace's medium supplemented with yeastolate, lactalbumin hydrolysate, and/or fetal calf serum as necessary (Gibco Life Technologies, Gaithersburg, MD).

Typically, an antibiotic or other compound useful for selective growth of transfected or transformed cells is added as a supplement to the media. The compound to be used will be dictated by the selectable marker element present on the plasmid with

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which the host cell was transformed. For example, where the selectable marker element is kanamycin resistance, the compound added to the culture medium will be kanamycin; where the selectable marker element is ampicillin resistance, the compound added to the culture medium will be ampicillin.

The amount of an AGP-1 polypeptide produced by a host cell can be evaluated using standard methods known in the art. Such methods include, without limitation, Western blot analysis, SDS-polyacrylamide gel electrophoresis, non-denaturing gel electrophoresis, HPLC separation, immunoprecipitation, and/or activity assays such as DNA binding gel shift assays.

Where an AGP-1 polypeptide is prepared without a tag attached, and no antibodies are available, other well known procedures for purification can be used. Such procedures include, without limitation, ion exchange chromatography, molecular sieve chromatography, HPLC, native gel electrophoresis in combination with gel elution, and preparative isoelectric focusing ("Isoprime" machine/technique, Hoefer Scientific). In some cases, two or more of these techniques may be combined to achieve increased purity.

If an AGP-1 polypeptide is produced intracellularly, the intracellular material (including inclusion bodies for gram-negative bacteria) can be extracted from the host cell using any standard technique known to the skilled artisan. For example, the host cells can be lysed to release the contents of the periplasm/cytoplasm by French press, homogenization, and/or sonication followed by centrifugation.

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35 If an AGP-1 polypeptide has formed inclusion bodies in the cytosol, the inclusion bodies can often

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bind to the inner and/or outer cellular membranes and thus will be found primarily in the pellet material after centrifugation. The pellet material can then be treated at pH extremes or with chaotropic agent such as a detergent, guanidine, guanidine derivatives, urea, or urea derivatives in the presence of a reducing agent such as dithiothreitol at alkaline pH or tris carboxyethyl phosphine at acid pH to release, break apart, and solubilize the inclusion bodies. An AGP-1 polypeptide in its now soluble form can then be analyzed using gel electrophoresis, immunoprecipitation or the like. If it is desired to isolate an AGP-1 polypeptide, isolation may be accomplished using standard methods such as those set forth below and in Marston et al. (Meth. Enz., 182, 264-275 (1990)).

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In some cases, an AGP-1 polypeptide may not be biologically active upon isolation. Various methods for "refolding" or converting the polypeptide to its tertiary structure and generating disulfide linkages, can be used to restore biological activity. 20 methods include exposing the solubilized polypeptide to a pH usually above 7 and in the presence of a particular concentration of a chaotrope. The selection of chaotrope is very similar to the choices used for inclusion body solubilization, but usually the 25 chaotrope is used at a lower concentration and is not necessarily the same as chaotropes used for the solubilization. In most cases the refolding/oxidation solution will also contain a reducing agent or the reducing agent plus its oxidized form in a specific 30 ratio to generate a particular redox potential allowing for disulfide shuffling to occur in the formation of the protein's cysteine bridge(s). Some of the commonly used redox couples include cysteine/cystamine,

35 glutathione (GSH)/dithiobis GSH, cupric chloride,

dithiothreitol(DTT)/dithiane DTT, and 2-

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mercaptoethanol(bME)/dithio-b(ME). In many instances, a cosolvent may be used or may be needed to increase the efficiency of the refolding and the more common reagents used for this purpose include glycerol, polyethylene glycol of various molecular weights, arginine and the like.

## <u>Derivatives</u>

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The present Fc-AGP-1 fusion proteins, and 10 variants and fragments thereof, are derivatized by the attachment of one or more chemical moieties to an Fc-AGP-1 fusion protein moiety. These chemically modified derivatives may be further formulated for intraarterial, intraperitoneal, intramuscular subcutaneous, intravenous, oral, nasal, pulmonary, 15 topical or other routes of administration as discussed below. Chemical modification of biologically active proteins has been found to provide additional advantages under certain circumstances, such as increasing the stability and circulation time of the 20 therapeutic protein and decreasing immunogenicity. See, U.S. Patent No. 4,179,337. For a review, see Abuchowski et al., in Enzymes as Drugs. (J. S. Holcerberg and J. Roberts, eds. pp. 367-383 (1981)); 25 Francis et al., supra.

The chemical moieties suitable for such derivatization may be selected from among various water soluble polymers. The polymer selected should be water soluble so that the protein to which it is attached does not precipitate in an aqueous environment, such as a physiological environment. Preferably, for therapeutic use of the end-product preparation, the polymer will be pharmaceutically acceptable. One skilled in the art will be able to select the desired polymer based on such considerations as whether the polymer/protein conjugate will be used therapeutically,

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and if so, the desired dosage, circulation time, resistance to proteolysis, and other considerations. For the present proteins, the effectiveness of the derivatization may be ascertained by administering the derivative, in the desired form (i.e., by osmotic pump, or, more preferably, by injection or infusion, or, further formulated for oral, pulmonary or nasal delivery, for example), and observing biological effects as described herein.

10 The water soluble polymer may be selected from the group consisting of, for example, polyethylene glycol, copolymers of ethylene glycol/propylene glycol, carboxymethylcellulose, dextran, polyvinyl alcohol, polyvinyl pyrolidone, poly-1, 3-dioxolane,

poly-1,3,6-trioxane, ethylene/maleic anhydride
copolymer, polyaminoacids (either homopolymers or
random copolymers), and dextran or poly(n-vinyl
pyrolidone)polyethylene glycol, propylene glycol
homopolymers, polypropylene oxide/ethylene oxide
co-polymers, polyoxyethylated polyols and polyving

co-polymers, polyoxyethylated polyols and polyvinyl alcohol. Polyethylene glycol propionaldenhyde may have advantages in manufacturing due to its stability in water. Also, succinate and styrene may also be used.

Fc-AGP-1 fusion proteins, variants and
fragments may also be derivatized by attaching
polyaminoacids or branch point amino acids to the Fc or
AGP-1 protein (or variant or fragment) moiety. For
example, the polyaminoacid may be an additional carrier
protein which serves to further increase the
circulation half life of an Fc-AGP-1 fusion protein in

circulation half life of an Fc-AGP-1 fusion protein in addition to the advantages achieved via the Fc-AGP-1 fusion protein above. For the present therapeutic or cosmetic purpose of the present invention, such polyaminoacids should be those which have or do not

35 create neutralizing antigenic response, or other adverse responses. Such polyaminoacids may be selected

from the group consisting of serum album (such as human serum albumin), or other polyaminoacids, e.g. lysines. As indicated below, the location of attachment of the polyaminoacid may be at the N-terminus of the Fc-AGP-1 protein moiety, or C-terminus, or other places in between, and also may be connected by a chemical "linker" moiety to the Fc-AGP-1 protein.

The polymer may be of any molecular weight, and may be branched or unbranched. For polyethylene glycol, the preferred molecular weight is between about 10 2 kDa and about 100 kDa (the term "about" indicating that in preparations of polyethylene glycol, some molecules will weigh more, some less, than the stated molecular weight) for ease in handling and manufacturing. Other sizes may be used, depending on 15 the desired therapeutic profile (e.g., the duration of sustained release desired, the effects, if any on biological activity, the ease in handling, the degree or lack of antigenicity and other known effects of the polyethylene glycol to a therapeutic protein or 20 analog).

The number of polymer molecules so attached may vary, and one skilled in the art will be able to ascertain the effect on function. One may mono-derivatize, or may provide for a di-, tri-, tetra-25 or some combination of derivatization, with the same or different chemical moieties (e.g., polymers, such as different weights of polyethylene glycols). proportion of polymer molecules to protein (or peptide) molecules will vary, as will their concentrations in 30 the reaction mixture. In general, the optimum ratio (in terms of efficiency of reaction in that there is no excess unreacted protein or polymer) will be determined by factors such as the desired degree of derivatization (e.g., mono, di-, tri-, etc.), the molecular weight of 35

the polymer selected, whether the polymer is branched or unbranched, and the reaction conditions.

The chemical moieties should be attached to the protein with consideration of effects on functional or antigenic domains of the protein. There are a number of attachment methods available to those skilled in the art. E.g., EP 0401384 herein incorporated by reference (coupling PEG to G-CSF), see also Malik et al., Exp. Hematol. 20, 1028-1035 (1992) (reporting pegylation of GM-CSF using tresyl chloride). For 10 example, polyethylene glycol may be covalently bound through amino acid residues via a reactive group, such as, a free amino or carboxyl group. Reactive groups are those to which an activated polyethylene glycol molecule may be bound. The amino acid residues having 15 a free amino group may include lysine residues and the N-terminal amino acid residue. Those having a free carboxyl group may include aspartic acid residues, glutamic acid residues, and the C-terminal amino acid residue. Sulfhydryl groups may also be used as a 20 reactive group for attaching the polyethylene glycol molecule(s). Preferred for therapeutic purposes is attachment at an amino group, such as attachment at the N-terminus or lysine group. Attachment at residues important for receptor binding should be avoided if 25 receptor binding is desired.

One may specifically desire N-terminally chemically modified Fc-AGP-1 fusion protein. Using polyethylene glycol as an illustration of the present compositions, one may select from a variety of polyethylene glycol molecules (by molecular weight, branching, etc.), the proportion of polyethylene glycol molecules to protein (or peptide) molecules in the reaction mix, the type of pegylation reaction to be performed, and the method of obtaining the selected N-terminally pegylated protein. The method of

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obtaining the N-terminally pegylated preparation (i.e., separating this moiety from other monopegylated moieties if necessary) may be by purification of the N-terminally pegylated material from a population of pegylated protein molecules. Selective N-terminal chemical modification may be accomplished by reductive alkylation which exploits differential reactivity of different types of primary amino groups (lysine versus the N-terminal) available for derivatization in a particular protein. Under the appropriate reaction 10 conditions, substantially selective derivatization of the protein at the N-terminus with a carbonyl group containing polymer is achieved. For example, one may selectively N-terminally pegylate the protein by performing the reaction at a pH which allows one to 15 take advantage of the  $pK_a$  differences between the e-amino group of the lysine residues and that of the a-amino group of the N-terminal residue of the protein. By such selective derivatization, attachment of a water soluble polymer to a protein is controlled: the 20 conjugation with the polymer takes place predominantly at the N-terminus of the protein and no significant modification of other reactive groups, such as the lysine side chain amino groups, occurs. Using reductive alkylation, the water soluble polymer may be 25 of the type described above, and should have a single reactive aldehyde for coupling to the protein. Polyethylene glycol propionaldehyde, containing a single reactive aldehyde, may be used.

An N-terminally monopegylated derivative is preferred for ease in production of a therapeutic.

N-terminal pegylation ensures a homogenous product as characterization of the product is simplified relative to di-, tri- or other multi-pegylated products. The use of the above reductive alkylation process for

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preparation of an N-terminal product is preferred for ease in commercial manufacturing.

# Uses of the Polypeptide

AGP-1 fusion proteins may be used for the 5 treatment of proliferative disorders wherein cells are undergoing excessive proliferation. For example, an AGP-1 fusion protein may be used as an anti-tumor to treat patients suffering from a variety of cancers, such as breast cancer, prostate cancer, lung cancer, 10 and colon cancer. Viral infections and viral-induced diseases, such as hepatitis and AIDS, may also be treated with the proteins of the invention. Cardiovascular diseases such as arteriosclerosis which characterized by excessive proliferation of vascular 15 smooth muscle cells may also be treated. AGP-1 fusion proteins may also be used to suppress T-lymphocyte mediated immune responses that occur in autoimmune disorders and in rejection of transplanted tissues is desirable that that present proteins are expected to 20 have prolonged in vivo half-lives and circulation times that will allow for lower dosages, less frequent administration, and enhanced efficacy compared to an unfused AGP-1 protein.

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# Pharmaceutical Compositions

The present invention also provides for pharmaceutical compositions of the Fc-AGP-1 fusion proteins, variants, fragments and derivatives. Such 30 pharmaceutical compositions may be for administration for injection, or for oral, pulmonary, nasal, transdermal or other forms of administration. In general, comprehended by the invention are pharmaceutical compositions comprising effective 35 amounts of protein or derivative products of the invention together with pharmaceutically acceptable

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diluents, preservatives, solubilizers, emulsifiers, adjuvants and/or carriers. An effective or a therapeutically effective amount of an FcAGP-1 fusion protein is an amount sufficient to induce apoptosis in a target cell, wherein apoptosis is evaluated by assays known in the art.

Such compositions include diluents of various buffer content (e.g., Tris-HCl, acetate, phosphate), pH and ionic strength; additives such as detergents and 10 solubilizing agents (e.g., Tween 80, Polysorbate 80), anti-oxidants (e.g., ascorbic acid, sodium metabisulfite), preservatives (e.g., Thimersol, benzyl alcohol) and bulking substances (e.g., lactose, mannitol); incorporation of the material into 15 particulate preparations of polymeric compounds such as polylactic acid, polyglycolic acid, etc. or into liposomes. Hylauronic acid may also be used, and this may have the effect of promoting sustained duration in the circulation. Such compositions may influence the 20 physical state, stability, rate of in vivo release, and rate of in vivo clearance of the present proteins and derivatives. See, e.g., Remington's Pharmaceutical Sciences, 18th Ed. (1990, Mack Publishing Co., Easton, PA 18042) pages 1435-1712 which are herein incorporated 25 by reference. The compositions may be prepared in liquid form, or may be in dried powder, such as lyophilized form. Implantable sustained release formulations are also contemplated, as are transdermal formulations. 30

Contemplated for use herein are oral solid dosage forms, which are described generally in Remington's Pharmaceutical Sciences, 18th Ed. 1990 (Mack Publishing Co. Easton PA 18042) at Chapter 89, which is herein incorporated by reference. Solid dosage forms include tablets, capsules, pills, troches

or lozenges, cachets or pellets. Also, liposomal or proteinoid encapsulation may be used to formulate the present compositions (as, for example, proteinoid microspheres reported in U.S. Patent No. 4,925,673).

5 Liposomal encapsulation may be used and the liposomes may be derivatized with various polymers (e.g., U.S. Patent No. 5,013,556). A description of possible solid dosage forms for the therapeutic is given by Marshall, K. In: Modern Pharmaceutics Edited by G. S. Banker and C. T. Rhodes Chapter 10, 1979, herein incorporated by reference. In general, the formulation will include the Fc-AGP-1 fusion protein (or analog or derivative), and inert ingredients which allow for protection against the stomach environment, and release of the

biologically active material in the intestine.

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Also specifically contemplated are oral dosage forms of the above derivatized proteins. FcAGP-1 fusion protein may be chemically modified so that oral delivery of the derivative is efficacious. Generally, the chemical modification contemplated is the 20 attachment of at least one moiety to the protein (or peptide) molecule itself, where said moiety permits (a) inhibition of proteolysis; and (b) uptake into the blood stream from the stomach or intestine. Also desired is the increase in overall stability of the 25 protein and increase in circulation time in the body. Examples of such moieties include: Polyethylene glycol, copolymers of ethylene glycol and propylene glycol, carboxymethyl cellulose, dextran, polyvinyl alcohol, polyvinyl pyrrolidone and polyproline. 30 Abuchowski and Davis, Soluble Polymer-Enzyme Adducts. In: "Enzymes as Drugs", Hocenberg and RAGP-lerts, eds., Wiley-Interscience, New York, NY, (1981), pp 367-383; Newmark, et al., J. Appl. Biochem. 4: 185-189 (1982). Other polymers that could be used are poly-1,3-

dioxolane and poly-1,3,6-tioxocane. Preferred for

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pharmaceutical usage, as indicated above, are polyethylene glycol moieties.

To ensure resistance to degradation in the stomach following oral administration, a coating impermeable to at least pH 5.0 is essential. Examples of the more common inert ingredients that are used as enteric coatings for oral formulations are cellulose acetate trimellitate (CAT),

hydroxypropylmethylcellulose phthalate (HPMCP), HPMCP 50, HPMCP 55, polyvinyl acetate phthalate (PVAP), 10 Eudragit L30D, Aquateric, cellulose acetate phthalate (CAP), Eudragit L, Eudragit S, and Shellac. These coatings may be used as mixed films.

The therapeutic can be included in the formulation as fine multiparticulates in the form of 15 granules or pellets of particle size about 1 mm. formulation of the material for capsule administration could also be as a powder, lightly compressed plugs or even as tablets. The therapeutic could be prepared by 20 compression.

One may dilute or increase the volume of an FcAGP-1 composition with an inert material. diluents could include carbohydrates, especially mannitol, α-lactose, anhydrous lactose, cellulose,

sucrose, modified dextrans and starch. Certain 25 inorganic salts may be also be used as fillers including calcium triphosphate, magnesium carbonate and sodium chloride. Some commercially available diluents are Fast-Flo, Emdex, STA-Rx 1500, Emcompress and Avicell.

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Disintegrants may be included in the formulation of the therapeutic into a solid dosage form. Materials used as disintegrates include but are not limited to starch including the commercial

disintegrant based on starch, Explotab. Sodium starch 35 glycolate, Amberlite, sodium carboxymethylcellulose,

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ultramylopectin, sodium alginate, gelatin, orange peel, acid carboxymethyl cellulose, natural sponge and bentonite may all be used. Another form of the disintegrants are the insoluble cationic exchange resins. Powdered gums may be used as disintegrants and as binders and these can include powdered gums such as agar, Karaya or tragacanth. Alginic acid and its sodium salt are also useful as disintegrants.

Binders may be used to hold the therapeutic

agent together to form a hard tablet and include
materials from natural products such as acacia,
tragacanth, starch and gelatin. Others include methyl
cellulose (MC), ethyl cellulose (EC) and carboxymethyl
cellulose (CMC). Polyvinyl pyrrolidone (PVP) and
hydroxypropylmethyl cellulose (HPMC) could both be used
in alcoholic solutions to granulate the therapeutic.

An antifrictional agent may be included in the formulation of the therapeutic to prevent sticking during the formulation process. Lubricants may be used as a layer between the therapeutic and the die wall, and these can include but are not limited to; stearic acid including its magnesium and calcium salts, polytetrafluoroethylene (PTFE), liquid paraffin, vegetable oils and waxes. Soluble lubricants may also be used such as sodium lauryl sulfate, magnesium lauryl sulfate, polyethylene glycol of various molecular weights, Carbowax 4000 and 6000.

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Glidants that might improve the flow properties of the drug during formulation and to aid rearrangement during compression might be added. The glidants may include starch, talc, pyrogenic silica and hydrated silicoaluminate.

To aid dissolution of the therapeutic into the aqueous environment a surfactant might be added as a wetting agent. Surfactants may include anionic detergents such as sodium lauryl sulfate, dioctyl

sodium sulfosuccinate and dioctyl sodium sulfonate. Cationic detergents might be used and could include benzalkonium chloride or benzethomium chloride. The list of potential nonionic detergents that could be included in the formulation as surfactants are lauromacrogol 400, polyoxyl 40 stearate, polyoxyethylene hydrogenated castor oil 10, 50 and 60, glycerol monostearate, polysorbate 40, 60, 65 and 80, sucrose fatty acid ester, methyl cellulose and carboxymethyl cellulose. These surfactants could be present in the formulation of the protein or derivative either alone or as a mixture in different ratios.

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Additives which potentially enhance uptake of the protein (or derivative) are for instance the fatty acids oleic acid, linoleic acid and linolenic acid.

Controlled release formulation may be desirable. The drug could be incorporated into an inert matrix which permits release by either diffusion or leaching mechanisms e.g., gums. Slowly degenerating matrices may also be incorporated into the formulation, e.g., alginates, polysaccahrides. Another form of a controlled release of this therapeutic is by a method based on the Oros therapeutic system (Alza Corp.), i.e., the drug is enclosed in a semipermeable membrane which allows water to enter and push drug out through a single small opening due to osmotic effects. Some enteric coatings also have a delayed release effect.

Other coatings may be used for the formulation. These include a variety of sugars which could be applied in a coating pan. The therapeutic agent could also be given in a film coated tablet and the materials used in this instance are divided into 2 groups. The first are the nonenteric materials and include methyl cellulose, ethyl cellulose, hydroxyethyl cellulose, methylhydroxy-ethyl cellulose, hydroxypropyl cellulose, hydroxypropyl-methyl cellulose, sodium

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carboxy-methyl cellulose, providone and the polyethylene glycols. The second group consists of the enteric materials that are commonly esters of phthalic acid.

A mix of materials might be used to provide the optimum film coating. Film coating may be carried out in a pan coater or in a fluidized bed or by compression coating.

Also contemplated herein is pulmonary

delivery of the present protein (or derivatives
thereof). The protein (or derivative) is delivered to
the lungs of a mammal while inhaling and traverses
across the lung epithelial lining to the blood stream.
(Other reports of this include Adjei et al.,

- Pharmaceutical Research 7: 565-569 (1990); Adjei et al., International Journal of Pharmaceutics 63: 135-144 (1990) (leuprolide acetate); Braquet et al., Journal of Cardiovascular Pharmacology 13 (suppl. 5): s.143-146 (1989) (endothelin-1); Hubbard et al., Annals of Internal
- 20 Medicine 3: 206-212 (1989) (α1-antitrypsin); Smith et al., J. Clin. Invest. 84: 1145-1146 (1989) (α1-proteinase); Oswein et al., "Aerosolization of Proteins", Proceedings of Symposium on Respiratory Drug Delivery II, Keystone, Colorado, March, 1990
- (recombinant human growth hormone); Debs et al., The Journal of Immunology  $\underline{140}$ : 3482-3488 (1988) (interferon  $\gamma$  and tumor necrosis factor  $\alpha$ ) and U.S. Patent No. 5,284,656 (granulocyte colony stimulating factor).

Contemplated for use in the practice of this
invention are a wide range of mechanical devices
designed for pulmonary delivery of therapeutic
products, including but not limited to nebulizers,
metered dose inhalers, and powder inhalers, all of
which are familiar to those skilled in the art.

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Some specific examples of commercially available devices suitable for the practice of this invention are the Ultravent nebulizer, manufactured by Mallinckrodt, Inc., St. Louis, Missouri; the Acorn II nebulizer, manufactured by Marquest Medical Products, Englewood, Colorado; the Ventolin metered dose inhaler, manufactured by Glaxo Inc., Research Triangle Park, North Carolina; and the Spinhaler powder inhaler, manufactured by Fisons Corp., Bedford, Massachusetts.

All such devices require the use of formulations suitable for the dispensing of protein (or analog or derivative). Typically, each formulation is specific to the type of device employed and may involve the use of an appropriate propellant material, in addition to diluents, adjuvants and/or carriers useful in therapy.

The protein (or derivative) should most advantageously be prepared in particulate form with an average particle size of less than 10  $\mu m$  (or microns), most preferably 0.5 to 5  $\mu m$ , for most effective delivery to the distal lung.

Carriers include carbohydrates such as trehalose, mannitol, xylitol, sucrose, lactose, and sorbitol. Other ingredients for use in formulations may include DPPC, DOPE, DSPC and DOPC. Natural or synthetic surfactants may be used. Polyethylene glycol may be used (even apart from its use in derivatizing the protein or analog). Dextrans, such as cyclodextran, may be used. Bile salts and other related enhancers may be used. Cellulose and cellulose derivatives may be used. Amino acids may be used, such as use in a buffer formulation.

Also, the use of liposomes, microcapsules or microspheres, inclusion complexes, or other types of carriers is contemplated.

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Nasal delivery of the protein (or analog or derivative) is also contemplated. Nasal delivery allows the passage of the protein to the blood stream directly after administering the therapeutic product to the nose, without the necessity for deposition of the product in the lung. Formulations for nasal delivery include those with dextran or cyclodextran. Delivery via transport across other mucus membranes is also contemplated.

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#### Dosage

One skilled in the art will be able to ascertain effective dosages by administration and observing the desired therapeutic effect. Due to the modification of an AGP-1 protein by fusion to an Fc, the present invention provides unexpected protein protection from degradation, and increases circulation time and stability, when compared to a soluble AGP-1 protein. One skilled in the art, therefore, will be able to ascertain from these changes that an effective dosage may require lower doses or less frequent dosing.

Preferably, the formulation of the molecule will be such that between about .10 µg/kg and 10 mg/kg will yield the desired therapeutic effect. The frequency of administration may be readily determined by one skilled in the art. The effective dosages may be determined using diagnostic tools over time. For example, a diagnostic for measuring the amount of AGP-1 protein or Fc-AGP-1 fusion protein in the blood (or plasma or serum) may first be used to determine endogenous levels of protein. Such diagnostic tools may be in the form of an antibody assay, such as an antibody sandwich assay. The amount of endogenous AGP-1 protein is quantified initially, and a baseline is determined. The therapeutic dosages are determined as the quantification of endogenous and exogenous AGP-1

protein or Fc-AGP-1 fusion protein (that is, protein, variant, fragment or derivative found within the body, either self-produced or administered) is continued over the course of therapy. The dosages may therefore vary over the course of therapy, with a relatively high dosage being used initially, until therapeutic benefit is seen, and lower dosages used to maintain the therapeutic benefits.

The following examples are offered to more fully illustrate the invention, but are not construed as limiting the scope thereof.

#### 15 EXAMPLE 1

## Production of AGP-1 Fusion Proteins

The fusion proteins Fc-huAGP-1 (95-281), Fc-huAGP-1 (114-281), Fc-muAGP-1 (99-291) and Fc-muAGP-1 (120-291) were constructed by the following procedures.

The human IgGyl Fc region was PCR amplified with the following set of oligonucleotide primers:

- 5' TCT CCA AGC TTG AGC CCA AAT CTT GTG ACA AAA C 3'
  25 (SEQ ID NO. 1)
  5' TCT CCC TTA AGT TTA CCC GGA GAC AGG GAG AG 3' (SEQ ID NO. 2)
- PCR reaction was carried out in a volume of 50 μl with 1 unit of vent DNA polymerase (New England Biolabs) in 20 mM Tris-HCl pH8.8, 10 mM KCl, 10 mM (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, 0.1% Triton-X100, 10 μM of each dNTP, 1μM of each primer and 10 ng of RANKFC/pCEPP4 template (Hsu et al, Proc. Natl. Acad. Sci. USA 96, 3540-3545 (1999)). Reactions were performed in 94°C for 30 sec., 55°C for 30 sec., and

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72°C for 1 min, for a total of 16 cycles. The PCR fragment was isolated by electrophoresis through 1% agarose and purification by the Geneclean procedure (Bio 101, Inc.). The PCR fragment creates a HindIII restriction site at 5' end and a AflII restriction site at 3' end. The HindIII-AflII digested PCR fragment was then subcloned into the pCEP4 vector (Invitrogen) to create Fc/pCEP4.

A murine OPG signal peptide having the 10 following amino acid sequence:

MNKWLCCALLVLLDIIEWTTQ (SEQ ID NO. 3) was created by annealing the following set of oligonucleotides:

- 5' CTA GCA CCA TGA ACA AGT GGC TGT GCT GCG CAC TCC TGG TGC TCC TGG ACA TCA TTG AAT GGA CAA CCC AGA 3' (SEQ ID NO. 4)
- 5' AGC TTC TGG GTT GTC CAT TCA ATG ATG TCC AGG AGC 20 ACC AGG AGT GCG CAG CAC AGC CAC TTG TTC ATG GTG 3' (SEQ ID NO. 5)

The annealing was carried out by heating 10 µM of each oligonucleotide in a total volume of 20 µl at 94°C for 5 minutes, and then cooling gradually to room temperature. The annealed oligonucleotides, creating NheI restriction site overhang at 5' end and HindIII restriction site overhang at 3' end, were cloned into Fc-pCEP4 vector inframe N-terminal to the human IgGγ1 Fc region to generate SO-Fc/pCEP4 vector. A linker which encodes two irrelevant amino acids (QK) was introduced between the OPG signal peptide and human IgGγ1 Fc. An extra irrelevant amino acid (K) was introduced by the HindIII site between the OPG signal

peptide and human IgGyl Fc.

The baculovirus expression system chosen to express nucleic acid sequences encoding Fc-AGP-1 fusion proteins is based on the BAC-TO-BACK expression system

[Life Technologies, Gaithersburg, MD] with the following modifications.

The major capsid protein promoter sequence
(GenBank Acc. No. M22978) was PCR amplified from the

10 Bac-N-Blue™ linear AcMNPV DNA purchased from Invitrogen
(Carlsbad, CA) with the following primers:

- 5' ATT ATT GAT ATC GCA TGC TTG TTC GCC ATC GTG GAA TC (SEO ID NO. 6)
- 15 5' AAT CCG GAA TAT TGT TGC CGT TAT AAA TAT GGA C (SEQ ID NO. 7)

The N-terminal coding sequence of the first 12 codons of the polyhedrin gene with the methionine start codon mutated from ATG to ATT and the following MCS were PCR amplified from the vector pBlueBac4 DNA (Invitrogen) with the following primers:

- 5' AAC GGC AAC AAT ATT CCG GAT TAT TCA TAC CGT CC (SEQ 25 ID NO. 8)
  - 5' ACT TCA AGG AGA ATT TCC (SEQ ID NO. 9)

The resulting two fragments share overlapping sequence and a second round of PCR was performed to fuse them together. The polyhedrin promoter and the multiple cloning sites (MCS) between the SnaBI and HindIII site present in pFastBacl was then replaced with the EcoRV and HindIII digested fragment containing capsid promoter and the new MCS. The resulting vector is named pFC. To further modify the MCS, the following

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pair of primers were annealed together and ligated to the pFC vector digested with NheI and HindIII.

5' CTA GCT CTA GAC ATA TGG AAT TCC TGC AGC AGC TGG TAC

CTC GAG GATCCA AGC TTG TCG ACT(SEQ ID NO. 10)

5' AGC TAG TCG ACA AGC TTG GAT CCT CGA GGT ACC AGC TGC

TGC AGG AAT TCC ATA TGT CTA GAG (SEQ ID NO. 11)

The resulting vector is named pFC1 and is used as the donor vector for all baculovirus expression work described here.

To generate Fc-AGP-1 sequences in a baculovirus expression vector, the SO-Fc/pCEP4 was first used as template for PCR amplification of OPG signal peptide followed by human IgGyl Fc region with the following primers:

- 5' GGG CGT GCT AGC CAC CAT GAA CAA GTG GCT GTG CTG C 3'
  20 (SEQ ID NO. 12)
  5' AGC TCC TTC TGC AGG TGG AAC AGC TGT TTA CCC GGA GAC
  AGG GAG 3' (SEQ ID NO. 13)
- PCR reactions were carried out under conditions similar to those described above. The PCR fragment creates NheI restriction site at 5' end and PstI restriction site at 3' end, and was subsequently cloned into pFC1 vector to create SO-Fc/pFC1.
- Human AGP-1(114-281) was PCR amplified from human AGP-1 and human IgGyl cDNA templates by the following overlapping set of primers to generate PCR fusion fragments:
- 35 5' CTC CGG GTA AAG TGA GAG AAA GAG GTC CTC AG 3' (SEQ ID NO. 14)

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- 5' TTC TCT CAC TTT ACC CGG AGA CAG GGA G 3' (SEQ ID NO. 15)
- 5' CTT CTT CCT CTA CAG CAA GC 3' (SEQ ID NO. 16)
- 5' GTT ATT GCT CAG CGG TGG CA 3' (SEQ ID NO. 17)

5

Human AGP-1(95-281) was PCR amplified from human AGP-1 and human IgGyl cDNA templates by the following overlapping set of primers to generate PCR fusion fragments:

10

- 5' CCG GGT AAA ACT TCT GAG GAA ACC ATT TCT AC 3' (SEQ ID NO. 18)
- 5' TCC TCA GAA GTT TTA CCC GGA GAC AGG GAG AG 3' (SEQ ID NO. 19)
- 15 5' CTT CTT CCT CTA CAG CAA GC 3' (SEQ ID NO. 20)
  - 5' GTT ATT GCT CAG CGG TGG CA 3' (SEQ ID NO. 21)

Murine AGP-1(120-291) was PCR amplified from murine AGP-1 and human IgG g1 cDNA templates by the following overlapping set of primers to generate PCR fusion fragments:

- 5' CCG GGT AAA GGT GGA AGA CCT CAG AAA GTG 3' (SEQ ID NO. 22)
- 25 5' GAG GTC TTC CAC CTT TAC CCG GAG ACA GGG AG 3' (SEQ ID NO. 23)
  - 5' CTT CTT CCT CTA CAG CAA GC 3' (SEQ ID NO. 24)
  - 5' GTT ATT GCT CAG CGG TGG CA 3' (SEQ ID NO. 25)
- 30 Murine AGP-1(99-291) was PCR amplified from human AGP-1 and human IgG g1 cDNA templates by the following overlapping set of primers to generate PCR fusion fragments:
- 35 5' CTC CGG GTA AAA CCT TTC AGG ACA CCA TTT CTA C 3' (SEQ ID NO. 26)

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5' CCT GAA AGG TTT TAC CCG GAG ACA GGG AG 3' (SEQ ID NO. 27)

- 5' CTT CTT CCT CTA CAG CAA GC 3' (SEQ ID NO. 28)
- 5' GTT ATT GCT CAG CGG TGG CA 3' (SEQ ID NO. 29)

5

10

The PCR was carried out in similar conditions as described above. The resulting PCR fusion fragments were digested with NsiI and XhoI restriction enzymes and cloned in frame C-terminal to the human IgGgl Fc region in SO-Fc/pFCl at NsiI and XhoI sites. The final pFCl constructs expressing Fc-AGP-1 fusion proteins were subject to sequencing analysis.

Transformation of the donor plasmids, selection of the recombinant bacmids and isolation of the recombinant 15 bacmid DNAs followed the protocols provided by the manufacturer (Life Technologies). The recombinant bacmid DNAs described above were transfected into insect SF-9 cells by standard calcium chloride precipitation method to generate recombinant viruses using Grace's insect cell 20 medium with supplements (Invitrogen) and 10 % fetal bovine serum. The viruses were subsequently amplified using the same cell line and medium. The final titer of the viruses was estimated to be around 10° (pfu/ml) based on previous experiences. To produce Fc-AGP-1 fusion proteins, shake 25 flask expression was set up to infect insect Hi-Five cells at a density of 2 x 10° cells/ml grown in serum-free Ex-CELL405 medium from JRH Biosciences (Lenexa, KS). The amount of virus preparation used was about 1/10 of the total cell culture volume. Time course samples were collected and 30 analyzed by Western blot analysis in a pilot experiment and the harvest time was determined to be at 50-55 hours postinfection. Some product degradation was observed at later hours and when more cells ( > 10 %) started to show signs of death. The harvested conditioned medium was filter-35

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sterilized immediately and kept at  $-80^{\circ}\text{C}$  until purification step.

5 EXAMPLE 2

Purification of AGP-1 Fusion Proteins

Pharmacia Protein A Sepharose. The resin was

equilibrated with TBS containing 20mM Tris pH7.0 and
150mM NaCl before applying the media. Complete
protease inhibitor cocktail (Boehringer-Mannheim) was
added to the media according to the manufacturer's
instructions. The media was loaded, the column washed
with TBS, and protein was eluted using Gentle Elution
buffer (Pierce, Rockford, IL). Protein containing
fractions were pooled and submitted for in vitro
analysis.

20

#### EXAMPLE 3

#### Production of AGP-1 Protein

PCR amplification employing the following 25 primer pair and human AGP-1 cDNA template are used to generate various forms of human AGP-1.

5' ATT TGA TTC TAG AAG GAG GAA TAA CAT ATG GTT CGT GAA
CGT GGT CCA CAG CGT GTA GCA 3' (SEQ ID NO. 30)

30

5' TAT CCG CGG ATC CTC GAG TTA GCC AAC TAA AAA GGC CCC GAA 3' (SEQ ID NO. 31)

One primer introduces unique XbaI and NdeI

restrictions sites, an initiating Mmthionine codon, and optimized codons for the amino terminal protein of the

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gene. The other primer of the pair introduces a TAA stop codon, and a unique XhoI site following the carboxyl terminus of the gene. PCR and thermocycling was performed using standard recombinant DNA methodology. The PCR products were purified, restriction digested and inserted into the unique XbaI and XhoI sites of the vector pAMG21 (ATCC accession no. 98113), and transformed into the prototrophic E.coli strain 2596. The resulting construct pAMG21-huAGP-1 (114-281) was engineered to encode human AGP-1 from amino acids 114-281 (see Figure 2). After transformation, clones were selected, plasmid DNA was isolated and the sequence of the AGP-1 gene insert was confirmed.

10

Human AGP-1(114-281) was purified as follows. 15 40.5g of E. coli cell paste was homogenized in 20mM Tris, 10mM EDTA, pH7.5 with Complete protease inhibitors in a total volume of 250mL. Cells were lysed with two passes in a microfluidizer at70psi and then centrifuged at 14,000 rpm in a JA14 rotor for 60min. 20 The supernatant was filtered through  $0.45\mu m$  and  $0.22\mu m$ pore size filters. Aliquots were stored at -20°C. One 10mL aliquot was thawed and centrifuged. The supernatant was diluted 1:2 with 20mM Tris pH 7.5, adjusted to pH 7.5 then filtered through a  $0.45\mu m$ 25 filter. The sample was then applied to an 5mL SP Hi Trap column which had been equilibrated with the above buffer. The protein was eluted using a gradient from 0-0.5M NaCl in 20mM Tris pH 7.5. AGP-1 containing fractions were concentrated and then diafiltered into 30 PBS pH6.2 using a centriprep 30. The concentrated fraction was estimated to be 90% pure by SDS-PAGE stained with Coomassie Blue.

#### EXAMPLE 4

35 Biological Activity of AGP-1 and AGP-1 Fusion Proteins

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Jurkat cells (ATCC No. \_\_\_ ) were maintained in RPMI medium 1640 containing 10% fetal calf serum, 100 mg/ml penicillin G, and 100 mg/ml streptomycin (GIBCO). To study apoptotsis induced by FcAGP-1 recombinant protein, 250 ml of Jurkat cells (5 x 105 cells/ml) were seeded to each well of a 96 well plate. Cells were incubated in 5% CO2 at 37°C with indicated concentrations of hu AGP-1 (114-281), huFcAGP-1 (114-281), or human IgG (Sigma) for 24 hours. Alamar Blue (Biosource Inc.) was added to each well in an amount 10 equal to 10% of the culture volume. Cells were incubated for another 8 hours. Fluorescence was measured with excitation wavelength at 530nm and emission wavelength at 590nm in Bio-TEK FL500 Fluorescence Plate Reader (Bio-TEK Insturments Inc.). 15 Each experiment was performed in duplicates. The results shown in Figure 7 indicate enhanced apoptosis activity of FcAGP-1 compared to soluble AGP-1.

20 \* \* \*

While the present invention has been described in terms of preferred embodiments, it is understood that variations and modifications will occur to those skilled in the art. Therefore, it is intended that the appended claims cover all such equivalent variations which come within the scope of the invention as claimed.

#### WHAT IS CLAIMED IS:

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- A protein having a formula selected from the group consisting of: R<sub>1</sub> R<sub>2</sub> and R<sub>1</sub> L R<sub>2</sub>, wherein
   R<sub>1</sub> is a Fc protein, or variant or fragment thereof, R<sub>2</sub> is an AGP-1 protein, or variant or fragment thereof, and L is a linker.
- 2. The protein according to claim 1, where in the Fc protein is selected from the group consisting of:
  - (a) the Fc amino acid sequences as set forth in Figure 1;
  - (b) the amino acid sequence of subpart (a) having a different amino acid substituted or deleted in one or more of the following positions (using the numbering according to Figure 1):
    - (i) one or more cysteine residues;
    - (ii) one or more tyrosine residues;
- - (iv) leucine at position 20 deleted or substituted with glutamine;
  - (v) glutamic acid at position 103 deleted or substituted with an alanine;
  - (vi) lysine at position 105 deleted or substituted with an alanine;
  - (vii)lysine at position 107 deleted or substituted with an alanine;
- 30 (viii) deletion or substitution of one or more of the amino acids at positions 1, 2, 3, 4, and 5;
  - (ix) one or more residues substituted or deleted to ablate the Fc receptor binding site;

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	- 4 <i>/</i>
	(x) one or more residues substituted or
	deleted to ablate the complement (C1q) binding
	site; and
	<pre>(xi) a combination of subparts i-x;</pre>
5	<ul><li>(c) the amino acid sequence of subparts (a)</li></ul>
	or (b) having a methionyl residue at the
	N-terminus;
	(d) the Fc protein, or variant, fragment or
	derivative thereof, of any of subparts (a) through
10	(c) comprised of a chemical moiety connected to
	the protein moiety;
	(e) a derivative of subpart (d) wherein said
	chemical moiety is a water soluble polymer moiety;
	(f) a derivative of subpart (e) wherein said
15	water soluble polymer moiety is polyethylene
	glycol; and
	(g) a derivative of subpart (e) wherein said
	water soluble polymer moiety is attached at solely
	the N-terminus of said protein moiety.
20	
	3. The protein according to claim 1,
	wherein the AGP-1 protein, or variant, fragment or
	derivative thereof, is selected from the group
0.5	consisting of:
25	(a) the amino acid sequence X-281 wherein X is any residue from 95 to 114 inclusive as shown
	in Figure 2 (SEQ ID NO:34);
	(b) the amino acid sequence of subpart (a)
	having a methionyl residue at the N-terminus.
30	(c) the AGP-1 protein, or variant, fragment
50	or derivative thereof, of any of subparts (a) and
	(b) comprised of a chemical moiety connected to
	the protein moiety;

(d) a derivative of subpart (c) wherein said

chemical moiety is a water soluble polymer moiety;

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(e) a derivative of subpart (d) wherein said water soluble polymer moiety is polyethylene glycol;

- (f) A derivative of subpart (d) wherein said water soluble polymer moiety is a polyamino acid moiety; and
  - (g) a derivative of subpart (d) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.

10

5

4. The protein of claim 1 wherein the linker sequence is one or more amino acids selected from the group consisting of: Glycine, Asparagine, Serine, Threonine and Alanine.

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- 5. The protein of claim 1 wherein the linker is selected from the group consisting of:
  - (a) ala-ala-ala;
  - (b) ala-ala-ala;
- 20 (c) ala-ala-ala-ala;
  - (d) gly-gly;
  - (e) gly-gly-gly;
  - (f) gly-gly-gly-gly;
  - (g) gly-gly-gly-gly-gly-gly;
- 25 (h) gly-pro-gly;
  - (i) gly-gly-pro-gly-gly;
  - (j) chemical moiety; and
  - (k) any combination of subparts (a)

through (j).

30

6. A fusion protein comprising a Fc protein, or variant, fragment or derivative thereof, fused to the N-terminus of an AGP-1 protein, or variant, fragment or derivative thereof.

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- 7. A nucleic acid sequence encoding for a protein having the formula selected from the group consisting of:  $R_1 R_2$  and  $R_1 L R_2$ , wherein  $R_1$  is a Fc protein, or variant or fragment thereof,  $R_2$  is an AGP-1 protein, or variant or fragment thereof, and L is a linker.
- 8. The nucleic acid sequence according to claim 7 encoding for a protein comprising an Fc
  10 protein, variant, fragment or derivative portion selected from the group consisting of:
  - (a) the Fc amino acid sequences as set forth
    in Figure 1 (SEQ ID NO: 32);
  - (b) the amino acid sequence of subpart (a) having a different amino acid substituted or deleted in one or more of the following positions (using the numbering according to Figure 1):
    - (i) one or more cysteine residues;
    - (ii) one or more tyrosine residues;
    - (iii) cysteine at position 5 deleted or substituted with an alanine;
    - (iv) leucine at position 20 deleted or substituted with glutamine;
    - (v) glutamic acid at position 103deleted or substituted with an alanine;
    - (vi) lysine at position 105 deleted or substituted with an alanine;
    - (vii)lysine at position 107 deleted or substituted with an alanine;
    - (viii) deletion or substitution of one or more of the amino acids at positions 1, 2, 3, 4, and 5;
    - (ix) one or more residues substituted or deleted to ablate the Fc receptor binding site;

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	- 50 <i>-</i>
	(x) one or more residues substituted or
	deleted to ablate the complement (Clq) binding
	site; and
	<pre>(xi) a combination of subparts i-x;</pre>
5	(c) the amino acid sequence of subparts (a)
	or (b) having a methionyl residue at the
	N-terminus;
	(d) the Fc protein, or variant, fragment or
	derivative thereof, of any of subparts (a) through
10	(c) comprised of a chemical moiety connected to
	the protein moiety;
	(e) a derivative of subpart (d) wherein said
	chemical moiety is a water soluble polymer moiety;
	(f) a derivative of subpart (e) wherein said
15	water soluble polymer moiety is polyethylene
	glycol; and
	(g) a derivative of subpart (e) wherein said
	water soluble polymer moiety is attached at solely
	the N-terminus of said protein moiety.
20	
	9. The nucleic acid sequence according to
	claim 7 encoding for a protein comprising an AGP-1
	protein, variant, fragment or derivative portion
	selected from the group consisting of:
25	(a) the amino acid sequence X-281 wherein X
	is any residue from 95 to 114 inclusive as shown
	in Figure 2 (SEQ ID NO: 34);
	(b) the amino acid sequence of subpart (a)
	having a methionyl residue at the N-terminus.
30	(c) the AGP-1 protein, or variant, fragment
	or derivative thereof, of any of subparts (a) and
	(b) comprised of a chemical moiety connected to

(d) a derivative of subpart (c) wherein said

chemical moiety is a water soluble polymer moiety;

the protein moiety;

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	(e)	a	đe	rivat	ive	of	suk	par	t	(d)	wherein	said
water	sol	ubl	e	polym	er 1	noie	ety	is	oq	lyet	hylene	
glyco	1;											

- (f) A derivative of subpart (d) wherein said water soluble polymer moiety is a polyamino acid moiety; and
- (g) a derivative of subpart (d) wherein said water soluble polymer moiety is attached at solely the N-terminus of said protein moiety.

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10. The nucleic acid sequence of claim 7 encoding for a protein with a linker sequence of one or more amino acids selected from the group consisting of: Gly, Asn, Ser, Thr and Ala.

- 11. The nucleic acid sequence of claim 7 encoding for a protein with a linker selected from the group consisting of:
  - (a) ala, ala, ala;
- 20 (b) ala-ala-ala-ala;
  - (c) ala-ala-ala-ala;
  - (d) gly-gly;
  - (e) gly-gly-gly;
  - (f) gly-gly-gly-gly;
- 25 (g) gly-gly-gly-gly-gly-gly;
  - (h) gly-pro-gly;
  - (i) gly-gly-pro-gly-gly;
  - (j) a chemical moiety; and
  - (k) any combination of subparts (a)
- 30 through (j).
- 12. A nucleic acid sequence encoding for a fusion protein having a Fc protein, or variant, fragment or derivative thereof, fused to the N-terminus of an AGP-1 protein, or a variant, fragment or derivative thereof.

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- 13. A vector comprising a nucleic acid sequence according to any of Claims 7 to 12 inclusive.
- 5 14. A prokaryotic or eukaryotic host cell containing the vector of claim 13.
- 15. A process for producing a protein of claims 1 or 6 comprising the steps of culturing, under suitable conditions, the host cell of claim 14, and isolating the protein produced.
  - 16. The process of claim 15 further comprising the step of purifying the protein produced.

15

17. A pharmaceutical composition comprising an effective amount of a protein according to claims 1 or 6, in a pharmaceutically acceptable diluent, adjuvant or carrier.

20

18. A method of inducing apoptosis in a tissue comprising administering a therapeutically effective amount of the protein according to Claim 1 or 6.

25

19. A method of treating of a disorder selected from the group consisting of proliferative disorder, an immune disorder or a viral-induced disorder comprising administering a therapeutically effective amount of the protein according to claims 1 or 6.

# FIGURE 1 Amino acid sequence of hinge, CH2 and CH3 regions human IgGγ1

Glu 1	Pro	Lys	Ser	Суs 5	Asp	Lys	Thr	His	Thr 10	Cys	Pro	Pro	Cys	Pro 15	Ala
Pro	Glu	Leu	Leu 20	Gly	Gly	Pro	Ser	Val 25	Phe	Leu	Phe	Pro	Pro 30	Ŀys	Pro
Lys	qaA	Thr 35	Leu	Met	Ile	Ser	Arg 40	Thr	Pro	Glu	Va1	Thr 45	Cys	Val	Val
Val	Asp 50	Val	Ser	His	Glu	Asp 55	Pro	Glu	Val	Lys	Phe 60	Asn	Trp	Tyr	Val
Asp 65	Gly	Val	Glu	Val	His 70	Asn	Ala	Lys	Thr	Lys 75	Pro	Arg	Glu	Glu	Gln 80
Tyr	Asn	Ser	Thr	Tyr 85	Arg	Val	Val	Ser	Val 90	Leu	Thr	Val	Leu	His 95	Gln
Asp	Trp	Leu	Asn 100	Gly	Lys	Glu	Tyr	Lys 105	Cys	Lys	Val	Ser	Asn 110	Lys	Ala
Leu	Pro	Ala 115	Pro	Ile	Glu	Lys	Thr 120	Ile	Ser	Lys	Ala	Lys 125	Gly	Gln	Pro
Arg	Glu 130	Pro	Gln	Val	Tyr	Thr 135	Leu	Pro	Pro	Ser	Arg 140	Asp	Glu	Leu	Thr
Lys 145	Asn	Gln	Val	Ser	Leu 150	Thr	Cys	Leu	Val	Lys 155	Gly	Phe	Tyr	Pro	Ser 160
Asp	Ile	Ala	Val	Glu 165	Trp	Glu	Ser	Asn	Gly 170	Gln	Pro	Glu	Asn	Asn 175	Tyr
Lys	Thr	Thr	Pro 180	Pro	Val	Leu	Asp	Ser 185	Asp	Gly	Ser	Phe	Phe 190	Leu	Tyr
Ser	Lys	Leu 195	Thr	Val	Asp	Lys	Ser 200	Arg	Trp	Gln	Gln	Gly 205	Asn	Val	Phe
Ser	Cys 210	Ser	Val	Met	His	Glu 215	Ala	Leu	His	Asn	His 220	Tyr	Thr	Gln	Lys
Ser 225	Leu	Ser	Leu	Ser	Pro 230	Gly	Lys								

# FIGURE 2 Amino acid and nucleic acid sequence of Fc-huAGP-1 (95-281)

GGCTGACTTA CAGCAG	TCAG ACTCTGACA	G GATC ATG G	CT ATG ATG GAG GTG la Met Met Glu Val	C 52
CAG GGG GGA CCC A Gln Gly Gly Pro S 10	GC CTG GGA CAG er Leu Gly Gln	ACC TGC GTG Thr Cys Val 15	CTG ATC GTG ATC Leu Ile Val Ile 20	TTC 100 Phe
ACA GTG CTC CTG C Thr Val Leu Leu G 25	AG TCT CTC TGT ln Ser Leu Cys 30	: Val Ala Val	ACT TAC GTG TAC Thr Tyr Val Tyr	TTT 148 Phe
ACC AAC GAG CTG A Thr Asn Glu Leu L 40	AG CAG ATG CAG ys Gln Met Gln 45	GAC AAG TAC Asp Lys Tyr	TCC AAA AGT GGC . Ser Lys Ser Gly 50	ATT 196 Ile
GCT TGT TTC TTA A Ala Cys Phe Leu L 55	AA GAA GAT GAC ys Glu Asp Asp 60	AGT TAT TGG Ser Tyr Trp 65	GAC CCC AAT GAC Asp Pro Asn Asp	GAA 244 Glu 70
GAG AGT ATG AAC A Glu Ser Met Asn S	GC CCC TGC TGG er Pro Cys Trp 75	CAA GTC AAG Gln Val Lys 80	TGG CAA CTC CGT Trp Gln Leu Arg 85	CAG 292 Gln
CTC GTT AGA AAG A Leu Val Arg Lys M 90	TG ATT TTG AGA et lle Leu Arg	A ACC TCT GAG Thr Ser Glu 95	GAA ACC ATT TCT Glu Thr Ile Ser 100	ACA 340 Thr
GTT CAA GAA AAG C Val Gln Glu Lys G 105	AA CAA AAT ATT In Gln Asn Ile 110	ser Pro Leu	GTG AGA GAA AGA Val Arg Glu Arg 115	GGT 388 Gly
CCT CAG AGA GTA G Pro Gln Arg Val A 120	CA GCT CAC ATA la Ala His Ile 125	A ACT GGG ACC Thr Gly Thr	AGA GGA AGA AGC Arg Gly Arg Ser 130	AAC 436 Asn
ACA TTG TCT TCT C Thr Leu Ser Ser F 135	CA AAC TCC AAG Pro Asn Ser Lys 140	G AAT GAA AAG S Asn Glu Lys 145	Ala Leu Gly Arg	AAA 484 Lys 150
ATA AAC TCC TGG G Ile Asn Ser Trp G 1	AA TCA TCA AGG lu Ser Ser Arg 55	G AGT GGG CAT J Ser Gly His 160	TCA TTC CTG AGC Ser Phe Leu Ser 165	AAC 532 Asn
TTG CAC TTG AGG A Leu His Leu Arg A 170	AT GGT GAA CTO Asn Gly Glu Lev	G GTC ATC CAT 1 Val Ile His 175	GAA AAA GGG TTT Glu Lys Gly Phe 180	TAC 580 Tyr
TAC ATC TAT TCC C Tyr Ile Tyr Ser C 185	AA ACA TAC TTT In Thr Tyr Phe 190	e Arg Phe Gln	GAG GAA ATA AAA Glu Glu Ile Lys 195	GAA 628 Glu
AAC ACA AAG AAC C Asn Thr Lys Asn A 200	GAC AAA CAA ATG Asp Lys Gln Met 205	G GTC CAA TAT t Val Gln Tyr	ATT TAC AAA TAC lle Tyr Lys Tyr 210	ACA 676 Thr
AGT TAT CCT GAC ( Ser Tyr Pro Asp I 215	CCT ATA TTG TTC Pro Ile Leu Leu 220	G ATG AAA AGT u Met Lys Ser 225	Ala Arg Asn Ser	TGT 724 Cys 230
Trp Ser Lys Asp 1	GCA GAA TAT GG Ala Glu Tyr Gl 235	A CTC TAT TCC y Leu Tyr Ser 240	ATC TAT CAA GGG Tile Tyr Gln Gly 245	GGA 772 Gly

# FIGURE 2 (con't)

ATA Ile	TTT Phe	GAG Glu	CTT Leu 250	AAG Lys	GAA Glu	AAT Asn	GAC Asp	AGA Arg 255	ATT Ile	TTT Phe	GTT Val	TCT Ser	GTA Val 260	ACA Thr	AAT Asn	820
GAG Glu	CAC His	TTG Leu 265	ATA Ile	GAC Asp	ATG Met	GAC Asp	CAT His 270	GAA Glu	GCC Ala	AGT Ser	TTT Phe	TTC Phe 275	GGG Gly	GCC Ala	TTT Phe	868
	GTT Val 280		TAA *	CTG2	ACCTO	GA A	\AGA?	)AAA	ec a <i>i</i>	)AATA	CCTC1	AA A	etga(	CTAT		920
TCAC	TTT.	rca (	GAT	GATA(	A C	ratga	\AGAT	r GT	rtca/	AAAA	ATC	rgac(	CAA 2	AACAI	AACAAA	980
CAG	AAAA	CAG A	AAAA	CAAAC	A A	ACCTO	TATO	G CA	ATCT(	SAGT	AGA(	GCAG(	CCA (	CAAC	TAAAAT	1040
<b>ጥርጥ</b>	ATACA	AAC 2	ACAC	CATG	'A											1060

# FIGURE 3 Fc-huAGP-1(95-281)

	10				*****	****	30	`	,				5	n		
	10							-				-u -u -u -u			O to MILE	
GCTAGCC	accAT( M	JAAC N	AA( K	GTG( W	GCT( L	GTG C	CTGC	:GC/ A	ACT(	CCT	GGT	GCT(	L D	ACAT	_A1.	E E
	70						90						110			
ATGGACA	ACCCA(	GAAC	CT.	rga(	GCC	CAA	ATC:	rTG'	TGA	CAA	AAC	TCA	CACAT	GCCC.	ACC(	GTG
	T O 130	<u>K</u>	L	E	P	K	150	C	ט	K	1	n	T C	_	r	C
CCCAGCA	നസ്ത്രമ	አ ር ጥር	יירייטיי		GGG)	ACC	GTC	AGT	CTT	CCT	CTT	CCC	CCCAA	AACC	CAA	GGA
P A	P E	L	L	G	G	P	S	٧	F	L	F	P	P K	Р	K	D
'	190						210								~~~	
CACCCTC	ATGAT( M I	CTCC	CCG	GAC(	CCC'	rg≱ E	GGT( V	CAC.	ATG( C	CGT V	GGT V	GGT' V	GGACG D V	TGAG S	CCA H	CGA E
	250	5	• `	-	•	_	270		-				290			
AGACCCT	GAGGT	CAAC	TT	CAA	CTG	GTA	ACGTY	3GA	CGG	CGT	GGA	GGT	GCATA	ATGC	CAA	GAC
	E V 310	K	F	N	W	Y	V 330	D	G	V	E	V	н N 350	A	K	T
AAAGCCG	-	701	~~ × /		ሮንክ አለ	~ * ^		ጉም ን	ara.	יייביייי	Car	രമവ	CGTCC	ጥሮልሮ	CGT	cer
	CGGGA R E	E E	Q Q	Y	N	S	T	Y	R	V	V	S	V L	'I'	V	L
	370			_			390						410			
GCACCAG			JAA'	rģG	CAA	GGZ	GTA	CAA	GTG	CAA	.GGT	CTC	CAACA N K	AAGC	CCT 1.	CCC
	D W 430	L	N	G	~	£	450	K	_	21	v		470			
AGCCCCC	ATCGA	GAA	AAC	CAT	CTC	CAZ	AAGC	CAA	AGG	GCA	.GCC	CCG	AGAAC	CACA	GGT	GTA
ÄΡ	I E	ĸ	T	I	S	K	A 510	K	G	Q	P	R	E F	, Q	V	Y
	490									~~			•		o com	-
CACCCTG T L	CCCCC.			GGA D	TGA E	GC. L	rgac T	CAA K	GAA. N	CCA Q	v V	CAG S	L T	, C	L	V V
	550						570						590	)		
CAAAGGC		TCC	CAG	CGA	CAT	CG	CCGT	GGA	GTG	GGA	GAG	CAA	TGGG	AGCC	GGA	GAA N
	F Y 610	P	S	D	Ι	A	630		W	Е	5	IN	G (	) -	E,	14
CAACTAC	יא א בארי	ሮልሮ	מממ	шСС	CCT	ימכיי	TGGA	CTC	:CGA	CGO	CTC	CTI	CTTCC	TCTA	CAG	CAA
	K T	T	P	P	v	L	D	S	D	G	S	F	F I	ı X	S	K
	670						690 •									
GCTCACC L T	GTGGA	CAA	GAG	CAG	GTG W	GC:	AGCA Q	GGC G	GAA N	.CGI	rcti F	CTC	ATGCT 2 D	rccgi 3 V	'GAT M	'GCA H
ш т	730		_	•	**	ws	750	•		•	-		770			
TGAGGCT	CTGCA	CAA	CCA	CTA	CAC	GC.	AGAA	GAG	CCI	CTC	cca	GTC	TCCG	GTA	AAC	TTC
E A	ь н 790	N	H	Ä	T	Q	K 810	S	L	S	L	S	P (	s K	[T	S
TGAGGAA			ma A		mas				አጠማ	. 74. 74. 2		الافتداءات	"ጥር <b>ር ር</b>	ግጥ አርሞ	rcac	BAGA
TGAGGAA E E		TTC S	TAC T	AG1 V	.TCA Q	AG. E	ĸ	Q	Q	N	I.	S	P )	L V	R	E
	850						870	•					891			
AAGAGGT		GAG	AGI	AGC	AGC	TC	ACAT	'AAC	TGC	GA(	CCA	JAG(	GAAGA	AGCAZ	ACA( T	CATT L
R G	P Q 910	ĸ	٧	A	A	н	930	1	C.	1	K	9	95	Õ .	-	
GTCTTCT	rccaaa	CTC	CAA	GAZ	\TGA	\AA	AGGC	TC	rggo	GCC(	GCA	AAA!	raaac'	TCCT(	ggg <i>i</i>	AATC
S S			K	N	E	K	A	L	G	R	K	I	Ŋ	s w	E	S

### FIGURE 3 (con't)

ATCAAGGAGTGGGCATTCATTCCTGAGCAACTTGCACTTGAGGAATGGCGAACTGGTCAT
S R S G H S F L S N L H L R N G E L V I
1030 1050 1070

CCATGAAAAAAGGGTTTTACTACATCTATTCCCAAACATACTTTCGATTTCAGGAGGAAAT
H E K G F Y Y I Y S Q T Y F R F Q E E I
1090 1110 1130

AAAAGAAAACACAAAGAACGACAAACAAATGGTCCAATATATTTACAAATACACAAGTTA
K E N T K N D K Q M V Q Y I Y K Y T S Y
1150 1170 1190

TCCTGACCCTATATTGTTGATGAAAAGTGCTAGAAATAGTTGTTGGTCTAAAGATGCAGA
P D P I L L M K S A R N S C W S K D A E
1210 1230 1250

ATATGGACTCTATTCCATCTATCAAGGGGGGAATATTTGAGCATTAAGAATGACAGAAT
Y G L Y S I Y Q G G I F E L K E N D R I
1270 1290 1310

TTTTTGTTTCTGTAACAAATGAGCACTTGATAGGACATGACCATGAAGCCAGTTTTTTCGG
F V S V T N E H L I D M D H E A S F F G
1330

GGCCTTTTTAGTTGGCTAACCCGAG
GGCCTTTTTAGTTGGCTAACCCGAG
GGCCTTTTTAGTTGGCTAACCCCGAG
GGCCTTTTTAGTTGGCTAACCCCGAG
GGCCTTTTTAGTTGGCTAACCCCGAG
GGCCTTTTTAGTTGGCTAACCCCGAG
GGCCTTTTTAGTTGGCTAACCCCGAG
GGCCTTTTTAGTTGGCTAACCCCGAG
GGCCTTTTTAGTTGGCTAACCCCGGG
A F L V G] \*

## FIGURE 4 Fc-huAGP-1(114-281)

		1.0							30						5	0			
GCT	AGC	cacc.	ATG	AA	CAA	GTG	GCT	GT(	GCTG(	CGC	ACTO	CT	GGT	GCT	CCTG	GAC	CATO	CATT	GA
		70	М	N	ĸ	W	<u>L</u>	<u>C</u>	90	A	<u>L</u> _	<u></u>	<u>v</u>	<u>L</u>	11	<u>D</u>	<u>I</u>	_I	E
	GAC.	AACC T 130	Q	AA K	GCT L	TGA E	.GCC P	CAI K	AATC S 150	rtg C	TGA( D	K K	AAC T	TCA H	CACA T 17	C	P P	ACC(	etë C
	AGC: A		E	L	CCT L	GGG G	GGG G	AC(	GTC S 210	AGT V	CTT( F	CCT L	ĊTT F	CCC P	CCCA P 23	K	ACC(	CAA( K	GA D
	CCT( L		Ι	CTC S	CCG R	GAC T	CCC P	TG/ E	AGGTO V 270	CAC T	ATG C	CGT V	GGT V	GGT V	GGAC D 29	V	BAGO S	CCAC H	GA E
AGA(	CCC' P	IGAG E 310	V	CAA K	GTT F	CAA N	W.CTG	GTI Y	ACGTO V 330	GGA D	CGG G	CGT V	'GGA E	GGT V	GCAT H 35	N	rgc( A	CAA(	GAC T
AAA K	GCC(		E	GA E	GCA Q	GTA Y	CAA N	CA( S	GCAC T 390	GTA Y	CCG' R	TGT V	· GGT V	CAG S	CGTC V 41	Ŀ	CAC(	CGT( V	CT L
GCA H	CCA Q	GGAC D 430	M <sub>1</sub>	CT L	GAA N	TGC G	CAA K	AGGZ E	AGTA Y 450	CAA K	GTG C	CAA K	GGT V	CTC S	CAAC N 47	K	AGC( A	CCT(	P P
AGC A	CCC P	CATC I 490	E		AAC T	CAT I	CTC S	CA K	AAGC A 510	K	AGG G	GCA Q	GCC P	CCG R	AGAA E 53	P	ACA Q	GGT( V	GTA Y
CAC	CCT L	GCCC P 550	₽	ATC S	CCG R	GG <i>I</i> D	ATG <i>P</i> E	r P	TGAC T 570	K	GAA N	CC# Q	V V	CAG S	CCTG L 59	т	CTG( C	CCT( L	GT V
CAA K	AGG G		Y	PCC	CAG S	CGZ D	CAT I	CG A	CCGT V 630	E	GTG W	GG# E	AGAG S	CAA N	TGGG G 65	Q	GCC P	GGA(	gaa N
	CTA Y		T	CAC T	GCC P	TC( P	V V	rgc'	TGGA D 690	S	CGA D	CGC G	CTC S	CTI F	CTTC F 71	L	CTA Y	CAG S	CAA K
		CGTG V 730	D				GT( W	GGC Q	AGCA Q 750	G	N	CG1 V	rctt F	CTC S	ATGC C 77	S	CGT V	GAT M	GCA H
TGA E	.GGC A	TCTG L 790	Н	CAA N	CC? H	ACTZ Y	ACA( T	CGC Q	AGAA K 810	S	CCT L	'CT( S	ECC1	rgto S	P 83	G G O	TAA K	agt [V	GAG R
Ε	R	G 850	P )	Q	R	V	A	A	. н 870	)	T	G		R	GAGG <i>I</i> G 89	90 •	S	N	T.
Ļ	S	S 91(	P )	N	S	K	N	E	930	) )	L	G	R	K		N 50	S	W	# <u>.</u>
OTA	OTA:	AAGO	JAG	TGC	GC:	ATT S	CAT F	TCC L	TGAG	CAJ N	ACTI L	GC. H	ACT' L	TGA( R	GGAA: N	rgg G	CGA E	ACT L	'GGT V

# FIGURE 4 (con't)

		97	0						990						10	10			
CAT	CCA	TGA	AAA	AGG	TTT	CTA	CTA	CAT	'CTA'	TTC	CCA	AAC	ATA				TCA		
I	Н	E 103	К 0	G	F	Y	Y	1	.050	S	Q	Т	Y	F	R 10	F 70	Q	Ε	E
አ አጥ:	222	ACA	222	ሮልሮን	ነልል?	JAA:	CGA	CAA	ACA	AA7	rggt	CCA	ATA	TAT'	TTA	CAA	ATA	CAC	AAG
I	K	E 109	N	T	K	N	D	K	Q 110	M	V	Q	Y	I	Y 11	К 30	Y	T	S .
mas X i	ייייי	מ באיניי	୯୯୯	ጥልጥ	<u>ነ</u> ጥጥ	34147 •	GAT	GAA	AAG	TGO	TAG	AAA	TAG	TTG	TTG	GTC	TAA	AGA	TGC
Y	P	D 115	P	Ī	L	L	М	ĸ	.170	A	R	N	s	С	W 11	S	K	D	A
2.772	× 671.7	ATGG	* * CIE	ത്തു	nana.	יים איי	ימיזיי	מיייי	ACC	ccc	JAAT	ΑͲͲ	TGA	GCT	TAA	GGA	AAA'	TGA	CAG
E E	Ϋ́	G 121	L	Y	S	I	Y	Q	.230	G	I	F	E	L	K 12	E 50	N	D	R
* * **	mma	nenett	·	יותר ליוני	አአሮ፣		ጥርል	രവ	شىلىپ.	GAT	raga	САТ	GGA	CCA	TGA	AGC	CAG	TTT	TTT
I	F	V 127	S	V	T	N	E	H	L	I	D	M	D	Н	E	A	S	F	F
CGG	GGC	CTT	· TTT	AGT'	rgg(	CTA	Act	cga	ıg										
G	A.	F	L	v	G]	*		-	-										•

# FIGURE 5 Fc-muAGP-1(99-291)

20 1101101 2(1)	
10 30	50
GCTAGCCACCATGAACAAGTGGCTGTGCTGCGCACTCCTGG	TGCTCCTGGACATCATTGA
M N K W L C C A L L \	<u> </u>
70 90	ada da V
ATGGACAACCCAGAAGCTTGAGCCCAAATCTTGTGACAAA	ACTCACACATGCCCACCGTG
W T T O K L E P K S C D K 7	170
CCCAGCACCTGAACTCCTGGGGGGACCGTCAGTCTTCCTCTP A P E L L G G P S V F L E	PPKPKD
190 210	230
CACCCTCATGATCTCCCGGACCCCTGAGGTCACATGCGTGC	TGGTGGACGTGAGCCACGA
TLMISRTPEVTCVV	/ V D V S H E
250 270	290
AGACCCTGAGGTCAAGTTCAACTGGTACGTGGACGGCGTGC	BAGGTGCATAATGCCAAGAC
D P E V K F N W Y V D G V F 310 330	EVHNAKT 350
	· · · · · · · · · · · · · · · · · · ·
AAAGCCGCGGAGGAGCAGTACAACAGCACGTACCGTGTGC K P R E E Q Y N S T Y R V \	FTCAGCGTCCTCACCGTCCT
KPREEQYNSTYRV \ 370 390	410
GCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTGCAAG	מיירייירירים ארים א אבררריייריריר
H O D W L N G K E Y K C K	J S N K A L P
430 450	470
AGCCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAG	CCCGAGAACCACAGGTGTA
APIEKTISKAKGQI	PREPQVY
490 510	530
CACCCTGCCCCCATCCCGGGATGAGCTGACCAAGAACCAG	GTCAGCCTGACCTGCCTGGT
T L P P S R D E L T K N Q V 550 570	7 S L T C L V 590
CAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGA K G F Y P S D I A V E W E S	AGCAATGGGCAGCCGGAGAA S N G O P E N
KGFYPSDIAVEWE:	650
CAACTACAAGACCACGCCTCCCGTGCTGGACTCCGACGGC	<sup></sup> ₩ĊĊ₩₩Ċ₩₩ĊĊŴĊŢŖĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸĸ
N Y K T T P P V L D S D G	S F F L Y S K
670 690	710
GCTCACCGTGGACAAGAGCAGGTGGCAGCAGGGGAACGTC	TTCTCATGCTCCGTGATGCA
L T V D K S R W Q Q G N V 1	FSCSVMH
730 750	770
TGAGGCTCTGCACAACCACTACACGCAGAAGAGCCTCTCC	CTGTCTCCGGGTAAAacctt
EALHNHYTQKSLS 790 810	L S P G K [T F 830
tcaggacaccatttctacagttccagaaaagcagctaagt. ODTISTVPEKQLS	ACTCCTCCCTTGCCCAGAGG T P P L P R G
Q D T I S T V P E K Q L S ' 850 870	890
TGGAAGACCTCAGAAAGTGGCAGCTCACATTACTGGGATC	ACTOGGAGAAGCAACTCAGC
G R P Q K V A A H I T G I	TRRSNSA
910 930	950
TTTAATTCCAATCTCCAAGGATGGAAAGACCTTAGGCCAG	AAGATTGAATCCTGGGAGTC
LIPISKDGKTLGQ	KIESWES

#### FIGURE 5 (con't)

CTCTCGGAAAGGGCATTCATTTCTCAACCACGTGCTCTTTAGGAATGGAGAGCTGGTCAT
S R K G H S F L N H V L F R N G E L V I
1030 1050 1070

CGACCAGGAGGGCCTGTATTACATCTATTCCCAAACATACTTCCGATTTCAGGAAGCTGA
E Q E G L Y Y I Y S Q T Y F R F Q E A E
1090 1110 1130

AGACGCTTCAAGATGGTCTCAAAGGACAAGGTGAGACCAAACAGCTGGTGCAGTACAT
D A S K M V S K D K V R T K Q L V Q Y I
1150 1170 1190

CTACAAGTACACCAGCTATCCGGATCCCATAGTGCTCATGAAGAGCGCCAGAACAGCTG
Y K Y T S Y P D P I V L M K S A R N S C
1210 1230 1250

TTGGTCCAGAGATGCCGAGTACGGACTGTACTCCATCTATCAGGGAGGATTGTTCGAGCT
W S R D A E Y G L Y S I Y Q G G L F E L
1270 1290 1310

AAAAAAAAAATGACAGGATTTTTGTTTCTGTGACAAATGAACATTTGATGGACCTGGATCA
K K N D R I F V S V T N E H L M D L D Q
1330 1350

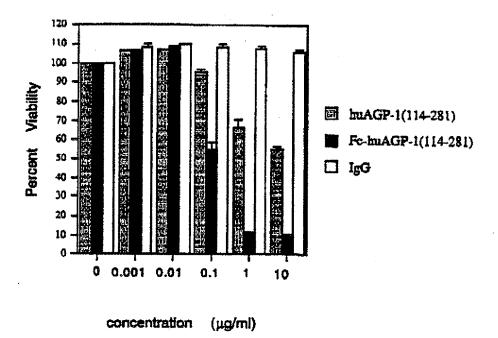
AGAAGCCAGCTTCTTTGGAGCCCTTTTTAATTAACTAACtaactcgag
E A S F F G A F L I N] \*

# FIGURE 6 Fc-muAGP-1(120-291)

	10			(A)			30						50			
GCTAGC	caccA]	ГGAA	CAA	GTG	GCT	GTO	GCTG(	CGC	ACT	ÇCT	GGT	GCT	CCTGG	ACAT	CATI	rga.
	70 70	N	<u>K</u>	W	L	<u>C</u>	<u>с</u> 90	<u>A</u>	<u>L</u>	<u>L</u>	V	<u>L</u>	<u>L D</u> 110	<u>I</u>	I	
ATGGAC W T							AATC' S 150		TGA D			TCA H	CACAT T C 170	GCCC P	ACCC P	TG C
CCCAGC P A	ACCTGA PE 190	AACT L			GGG G	AC(	CGTC S 210	agt V	CTT F	CCT L	CTT F	CCC P	CCCAA P K 230	AACC P	CAAC K	GA D
CACCCT T L	CATGAT M I 250		CCG R		CCC P	TG2 E	AGGTY V 270	CAC T	ATG C	CGT V	GGT V	GGT V	GGACG D V 290	TGAG S	CCAC H	GA E
AGACCC D P	TGAĞGT E V 310		GTT F				ACGTY V 330						GCATA H N 350		CAA( K	
AAAGCC K P	GCGGG/ R E 370	AGGA E	GCA Q	GTA Y	CAA N	CAC S	390	GTA Y	CCG R	TGT V	GGT V	CAG S	CGTCC V L 410	TCAC T	CGT( V	CCT L
GCACCA H Q	GGACTO D W 430	GCT L		TGG G	CAA K	.GG/ E	AGTA Y 450	CAA K	GTG C	CAA K	GGT V	CTC S	CAACA N K 470	A	CCT(	CCC P
AGCCCC A P	CATCGA I E 490		AAC T	CAT I	CTC S	CA.	AAGC A 510	CAA K	AGG G	GCA Q	GCC P	CCG R	AGAAC E P 530	CACA Q	GGT( V	STA Y
CACCCT T L	GCCCCC P P 550		CCG R				rgac T 570		GAA N	CCA Q	GGT V	CAG S	CCTGA L T 590	С	CCT(	GT V
CAAAGG K G	CTTCTA F Y 610		CAG S		CAT I	CG( A	CCGT V 630	GGA E	GTG W	GGA E	GAG S	CAA N	TGGGC G Q 650	P		GAA N
CAACTA N Y	CAAGAC K T 670		GCC ₽	TCC P	CGI V	'GC'	rgga D 690	S	CGA D	.CGG	CTC S	CTT F	CTTCC F L 710	Y	CAG( S	CAA K
GCTCAC L T	CGTGGI V D 730		GAG S			GC: Q		G	GAA N		CTI F	CTC S	ATGCT C S 770	V	GAT( M	GCA H
TGAGGC E A	TCTGCI L H 790	ACAA N	CCA H	CTA Y	CAC T	:GC: Q	AGAA K 810	S	ECT L	CTC S	CCT L	GTC S	TCCGG P G 830	K	AGG' [G	rgg G
	Q K 850	V	A	A	H	I	т 870	G	I	Т	TCO R	R	S N 890	S	A	L
	1 S 910	K	D	G	K	Ţ	930	G	Q	K	I	E	S W 950	E	S	s
TCGGAA	AGGGC	ATTC S	ATT F	TCT L	CAA N	ACC. H	ACGT V	GCT L	rcti F	TAC R	GAA N	ATGO G	AGAGC E I	TGGT V	rcat I	CGA E

### FIGURE 6 (con't)

# FIGURE 7



#### SEQUENCE LISTING

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<110> Amgen Inc.
<120> AGP-1 Fusion Protein Compositions and Methods
<130> A-600
<140> A-600
<141> 1999-04-16
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                                                                   32
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                                     10
Glu Trp Thr Thr Gln
             20
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ggacaaccca ga	72
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<220> <223> Description of Artificial Sequence: synthetic	
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ttgttcatgg tg	72
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<2,20>						
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<400> 10					<b>C</b> 0	
ctagctctag acatatggaa	ttectgcage aget	tggtac	c tegaggatee	aagcttgtcg	60	
act					63	
				•		
<210> 11						
<211> 62						
<212> DNA						
<213> Artificial Seque	nce					
<220>						
<223> Description of A	rtificial Seque	ence:	synthetic			
<400> 11					C0	
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gag					63	
<210> 12						
<211> 37						
<212> DNA						
<213> Artificial Seque	nce					
<220> <223> Description of A	rtificial Come	0000	evnthatic			
<2237 Description of A	remierar peda	C110 C +	ay ii cii cu c			
<400> 12	anaaraar ata	ctaa			37	
gggcgtgcta gccaccatga	acaageggee geg	uugu			**	
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F	
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.010. 00			
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Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val
Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val
Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln
                                                                                                                           75
                                                               70
Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln
                                                                                                              90
Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala
                                     100
Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro
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115

120

Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr 135 Lys Asn Gln Val Ser Leu Thr Cys Leu Val Lys Gly Phe Tyr Pro Ser 155 150 Asp Ile Ala Val Glu Trp Glu Ser Asn Gly Gln Pro Glu Asn Asn Tyr 170 Lys Thr Thr Pro Pro Val Leu Asp Ser Asp Gly Ser Phe Phe Leu Tyr 185 Ser Lys Leu Thr Val Asp Lys Ser Arg Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys 215 220 Ser Leu Ser Leu Ser Pro Gly Lys 230 <210> 33 <211> 1060 <212> DNA <213> Human <220> <221> CDS <222> Complement((35)..(877)) <400> 33 ggctgactta cagcagtcag actctgacag gate atg get atg gag gtc cag 55 Met Ala Met Met Glu Val Gln 103 ggg gga ccc agc ctg gga cag acc tgc gtg ctg atc gtg atc ttc aca Gly Gly Pro Ser Leu Gly Gln Thr Cys Val Leu Ile Val Ile Phe Thr gtg etc etg cag tet etc tgt gtg get gta aet tae gtg tae ttt aec Val Leu Leu Gln Ser Leu Cys Val Ala Val Thr Tyr Val Tyr Phe Thr aac gag ctg aag cag atg cag gac aag tac tcc aaa agt ggc att gct Asn Glu Leu Lys Gln Met Gln Asp Lys Tyr Ser Lys Ser Gly Ile Ala tgt ttc tta aaa gaa gat gac agt tat tgg gac ccc aat gac gaa gag Cys Phe Leu Lys Glu Asp Asp Ser Tyr Trp Asp Pro Asn Asp Glu Glu agt atg aac agc ccc tgc tgg caa gtc aag tgg caa ctc cgt cag ctc Ser Met Asn Ser Pro Cys Trp Gln Val Lys Trp Gln Leu Arg Gln Leu 80 gtt aga aag atg att ttg aga acc tct gag gaa acc att tct aca gtt Val Arg Lys Met Ile Leu Arg Thr Ser Glu Glu Thr Ile Ser Thr Val 1.00 90 95 caa gaa aag caa caa aat att tot ooc ota gtg aga gaa aga ggt oot

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Gln Glu Lys Gln Gln Asn Ile Ser Pro Leu Val Arg Glu Arg Gly Pro
                        110
cag aga gta gca gct cac ata act ggg acc aga gga aga agc aac aca
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Gln Arg Val Ala Ala His Ile Thr Gly Thr Arg Gly Arg Ser Asn Thr
                                        130
120
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Leu Ser Ser Pro Asn Ser Lys Asn Glu Lys Ala Leu Gly Arg Lys Ile
                                    145
aac tee tgg gaa tea tea agg agt ggg cat tea tte etg age aac ttg
Asn Ser Trp Glu Ser Ser Arg Ser Gly His Ser Phe Leu Ser Asn Leu
                                160
            155
cac ttg agg aat ggt gaa ctg gtc atc cat gaa aaa ggg ttt tac tac
                                                                   583
His Leu Arg Asn Gly Glu Leu Val Ile His Glu Lys Gly Phe Tyr Tyr
                            175
atc tat tcc caa aca tac ttt cga ttt cag gag gaa ata aaa gaa aac
                                                                   631
Ile Tyr Ser Gln Thr Tyr Phe Arg Phe Gln Glu Glu Ile Lys Glu Asn
    185
                        190
aca aag aac gac aaa caa atg gtc caa tat att tac aaa tac aca agt
                                                                   679
Thr Lys Asn Asp Lys Gln Met Val Gln Tyr Ile Tyr Lys Tyr Thr Ser
                    205
200
tat cct gac cct ata ttg ttg atg aaa agt gct aga aat agt tgt tgg
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Tyr Pro Asp Pro Ile Leu Leu Met Lys Ser Ala Arg Asn Ser Cys Trp
                220
tot aaa gat goa gaa tat gga oto tat too ato tat caa ggg gga ata
Ser Lys Asp Ala Glu Tyr Gly Leu Tyr Ser Ile Tyr Gln Gly Gly Ile
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            235
ttt gag ctt aag gaa aat gac aga att ttt gtt tct gta aca aat gag
                                                                   823
Phe Glu Leu Lys Glu Asn Asp Arg Ile Phe Val Ser Val Thr Asn Glu
                             255
cac ttg ata gac atg gac cat gaa gcc agt ttt ttc ggg gcc ttt tta
                                                                   871
His Leu Ile Asp Met Asp His Glu Ala Ser Phe Phe Gly Ala Phe Leu
                         270
gtt ggc taactgacct ggaaagaaaa agcaataacc tcaaagtgac tattcagttt
                                                                   927
Val Gly
280
tcaggatgat acactatgaa gatgtttcaa aaaatctgac caaaacaaac aaacagaaaa 987
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			225					230					235						
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ggt Gly	aaa Lys 255	act Thr	tct Ser	gag Glu	gaa Glu	acc Thr 260	att Ile	tct Ser	aca Thr	gtt Val	caa Gln 265	gaa Glu	aag Lys	caa Gln	caa Gln	817			
aat Asn 270	att Ile	tct Ser	ecc Pro	cta Leu	gtg Val 275	aga Arg	gaa Glu	aga Arg	ggt Gly	cct Pro 280	cag Gln	aga Arg	gta Val	gca Ala	gct Ala 285	865			
cac His	ata Ile	act Thr	ggg Gly	acc Thr 290	aga Arg	gga Gly	aga Arg	agc Ser	aac Asn 295	aca Thr	ttg Leu	tct Ser	tct Ser	cca Pro 300	aac Asn	913			
tcc Ser	aag Lys	aat Asn	gaa Glu 305	aag Lys	gct Ala	ctg Leu	ggc Gly	egc Arg 310	aaa Lys	ata Ile	aac Asn	tcc Ser	tgg Trp 315	gaa Glu	tca Ser	961			
tca Ser	agg Arg	agt Ser 320	Gly ggg	cat His	tca Ser	ttc Phe	ctg Leu 325	agc Ser	aac Asn	ttg Leu	cac His	ttg Leu 330	agg Arg	aat Asn	ggc	1009			
gaa Glu	ctg Leu 335	gtc Val	atc Ile	cat His	gaa Glu	aaa Lys 340	eja aaa	ttt Phe	tac Tyr	tac Tyr	ac atc tat tcc caa aca 1 yr Ile Tyr Ser Gln Thr 345 ac aca aag aac gac aaa 1 sn Thr Lys Asn Asp Lys								
tac Tyr 350	ttt Phe	cga Arg	ttt Phe	cag Gln	gag Glu 355	gaa Glu	ata Ile	aaa Lys	gaa Glu	aac Asn 360	aca Thr	aag Lys	aac Asn	gac Asp	aaa Lys 365	1105			
caa Gln	atg Met	gtc Val	caa Gln	tat Tyr 370	att Ile	tac Tyr	aaa Lys	tac Tyr	aca Thr 375	agt Ser	tat Tyr	cct Pro	gac Asp	cct Pro 380	ata Ile	1153			
ttg Leu	ttg Leu	atg Met	aaa Lys 385	agt Ser	gct Ala	aga Arg	aat Asn	agt Ser 390	tgt Cys	tgg Trp	tct Ser	aaa Lys	gat Asp 395	gca Ala	gaa Glu	1201			
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aat Asn	gac Asp 415	Arg	att Ile	ttt Phe	gtt Val	tct Ser 420	gta Val	aca Thr	aat Asn	gag Glu	cac His 425	Leu	ata Ile	gac	atg Met	1297			
gac Asp 430	His	gaa Glu	gcc Ala	agt Ser	ttt Phe 435	ttc Phe	ggg Gly	gcc Ala	ttt Phe	tta Leu 440	Val	ggc	taac	tcg	ag	1345			

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<212> PRT

<213> mouse

<400> 36

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- Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val
- Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr 50 55 60
- Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu 65 70 75 80
- Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys 85 90 95
- Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser 100 105 110
- Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys 115 120 125
- Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile 130 135 140
- Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro 145 150 155 160
- Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu 165 170 175
- Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn 180 185 190
- Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser 195 200 205
- Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg 210 215 220
- Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu 225 230 235 240
- His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys Thr 245 250 255
- Ser Glu Glu Thr Ile Ser Thr Val Gln Glu Lys Gln Gln Asn Ile Ser 260 265 270
- Pro Leu Val Arg Glu Arg Gly Pro Gln Arg Val Ala Ala His Ile Thr 275 280 285
- Gly Thr Arg Gly Arg Ser Asn Thr Leu Ser Ser Pro Asn Ser Lys Asn 290 295 300
- Glu Lys Ala Leu Gly Arg Lys Ile Asn Ser Trp Glu Ser Ser Arg Ser 305 310 315 320
- Gly His Ser Phe Leu Ser Asn Leu His Leu Arg Asn Gly Glu Leu Val

335 330 325 Ile His Glu Lys Gly Phe Tyr Tyr Ile Tyr Ser Gln Thr Tyr Phe Arg 345 340 Phe Gln Glu Glu Ile Lys Glu Asn Thr Lys Asn Asp Lys Gln Met Val Gln Tyr Ile Tyr Lys Tyr Thr Ser Tyr Pro Asp Pro Ile Leu Leu Met 370 Lys Ser Ala Arg Asn Ser Cys Trp Ser Lys Asp Ala Glu Tyr Gly Leu 395 385 Tyr Ser Ile Tyr Gln Gly Gly Ile Phe Glu Leu Lys Glu Asn Asp Arg 410 Ile Phe Val Ser Val Thr Asn Glu His Leu Ile Asp Met Asp His Glu 425 Ala Ser Phe Phe Gly Ala Phe Leu Val <210> 37 <211> 1288 <212> DNA <213> mouse <220> <221> CDS <222> Complement((11)..(1279)) <400> 37 getagecace atg aac aag tgg etg tge tge gea etc etg gtg etc etg Met Asn Lys Trp Leu Cys Cys Ala Leu Leu Val Leu Leu 5 gac atc att gaa tgg aca acc cag aag ctt gag ccc aaa tct tgt gac Asp Ile Ile Glu Trp Thr Thr Gln Lys Leu Glu Pro Lys Ser Cys Asp 20 aaa act cac aca tgc cca ccg tgc cca gca cct gaa ctc ctg ggg gga 145 Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly 35 30 ceg tea gto tto etc tto ecc eca aaa ecc aag gae acc etc atg atc 193 Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile tee egg ace eet gag gte aca tge gtg gtg gtg gae gtg age eac gaa 241 Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu 70 289 gac cct gag gtc aag ttc aac tgg tac gtg gac ggc gtg gag gtg cat Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His 85 aat goo aag aca aag cog ogg gag gag cag tac aac agc acg tac cgt

Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg

100

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gag Glu	tac Tyr	aag Lys	tgc Cys	aag Lys 130	gtc Val	tcc Ser	aac Asn	aaa Lys	gcc Ala 135	ctc Leu	cca Pro	gcc Ala	ccc Pro	atc Ile 140	gag Glu	433
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acc Thr	ctg Leu	ccc Pro 160	cca Pro	tcc Ser	cgg Arg	gat Asp	gag Glu 165	ctg Leu	acc Thr	aag Lys	aac Asn	cag Gln 170	gtc Val	agc Ser	ctg Leu	529
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ctg Leu	gac Asp	tcc Ser	gac Asp	ggc Gly 210	tcc Ser	ttc Phe	ttc Phe	ctc Leu	tac Tyr 215	agc Ser	aag Lys	ctc Leu	acc Thr	gtg Val 220	gac Asp	673
aag Lys	agc Ser	agg Arg	tgg Trp 225	cag Gln	cag Gln	GJÅ âāā	aac Asn	gtc Val 230	ttc Phe	tca Ser	tgc Cys	oc tcc gtg atg cat rs Ser Val Met His 235				721
											ctc tcc ctg tct ccg Leu Ser Leu Ser Pro					769
ggt Gly	aaa Lys 255	gtg Val	aga Arg	gaa Glu	aga Arg	ggt Gly 260	cct Pro	cag Gln	aga Arg	gta Val	gca Ala 265	gct Ala	cac His	ata Ile	act Thr	817
ggg Gly 270	acc Thr	aga Arg	gga Gly	aga Arg	agc Ser 275	Asn	aca Thr	Leu	Ser	Ser	Pro	aac Asn	tcc Ser	aag Lys	aat Asn 285	865
gaa Glu	aag Lys	gct Ala	ctg Leu	ggc Gly 290	cgc Arg	aaa Lys	ata Ile	aac Asn	tcc Ser 295	tgg Trp	gaa Glu	tca Ser	tca Ser	agg Arg 300	agt Ser	913
GJA aaa	cat His	tca Ser	ttc Phe 305	ctg Leu	agc Ser	aac Asn	ttg Leu	cac His 310	ttg Leu	agg Arg	aat Asn	ggc Gly	gaa Glu 315	ctg Leu	gtc Val	961
atc Ile	cat His	gaa Glu 320	aaa Lys	Gly aaa	ttt Phe	tac Tyr	tac Tyr 325	atc Ile	tat Tyr	tcc Ser	caa Gln	aca Thr 330	tac Tyr	ttt Phe	cga Arg	1009
ttt Phe	cag Gln 335	gag Glu	gaa Glu	ata Ile	aaa Lys	gaa Glu 340	aac Asn	aca Thr	aag Lys	aac Asn	gac Asp 345	aaa Lys	caa Gln	atg Met	gtc Val	1057

caa tat att tac aaa tac aca agt tat cct gac cct ata ttg ttg atg Gln Tyr Ile Tyr Lys Tyr Thr Ser Tyr Pro Asp Pro Ile Leu Leu Met 355 aaa agt gct aga aat agt tgt tgg tct aaa gat gca gaa tat gga ctc 1153 Lys Ser Ala Arg Asn Ser Cys Trp Ser Lys Asp Ala Glu Tyr Gly Leu tat tcc atc tat caa ggg gga ata ttt gag ctt aag gaa aat gac aga 1201 Tyr Ser Ile Tyr Gln Gly Gly Ile Phe Glu Leu Lys Glu Asn Asp Arg 390 att ttt gtt tct gta aca aat gag cac ttg ata gac atg gac cat gaa 1249 Ile Phe Val Ser Val Thr Asn Glu His Leu Ile Asp Met Asp His Glu 405 1288 gcc agt ttt ttc ggg gcc ttt tta gtt ggc taactcgag Ala Ser Phe Phe Gly Ala Phe Leu Val Gly 420 <210> 38 <211> 423 <212> PRT <213> mouse <400> 38 Met Asn Lys Trp Leu Cys Cys Ala Leu Leu Val Leu Leu Asp Ile Ile 10 5 Glu Trp Thr Thr Gln Lys Leu Glu Pro Lys Ser Cys Asp Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His Asn Ala Lys Thr Lys Pro Arg Glu Glu Gln Tyr Asn Ser Thr Tyr Arg Val Val Ser Val Leu Thr Val Leu His Gln Asp Trp Leu Asn Gly Lys Glu Tyr Lys Cys Lys Val Ser Asn Lys Ala Leu Pro Ala Pro Ile Glu Lys Thr Ile Ser Lys Ala Lys Gly Gln Pro Arg Glu Pro Gln Val Tyr Thr Leu Pro Pro Ser Arg Asp Glu Leu Thr Lys Asn Gln Val Ser Leu Thr Cys Leu 175 165 170 Val Lys Gly Phe Tyr Pro Ser Asp Ile Ala Val Glu Trp Glu Ser Asn

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WO 00/63253 185 190 Gly Gln Pro Glu Asn Asn Tyr Lys Thr Thr Pro Pro Val Leu Asp Ser 200 Asp Gly Ser Phe Phe Leu Tyr Ser Lys Leu Thr Val Asp Lys Ser Arg 215 Trp Gln Gln Gly Asn Val Phe Ser Cys Ser Val Met His Glu Ala Leu 230 His Asn His Tyr Thr Gln Lys Ser Leu Ser Leu Ser Pro Gly Lys Val 250 Arg Glu Arg Gly Pro Gln Arg Val Ala Ala His Ile Thr Gly Thr Arg 265 Gly Arg Ser Asn Thr Leu Ser Ser Pro Asn Ser Lys Asn Glu Lys Ala 280 Leu Gly Arg Lys Ile Asn Ser Trp Glu Ser Ser Arg Ser Gly His Ser 295 Phe Leu Ser Asn Leu His Leu Arg Asn Gly Glu Leu Val Ile His Glu 315 310 Lys Gly Phe Tyr Tyr Ile Tyr Ser Gln Thr Tyr Phe Arg Phe Gln Glu 330 Glu Ile Lys Glu Asn Thr Lys Asn Asp Lys Gln Met Val Gln Tyr Ile 345

Tyr Lys Tyr Thr Ser Tyr Pro Asp Pro Ile Leu Leu Met Lys Ser Ala

Arg Asn Ser Cys Trp Ser Lys Asp Ala Glu Tyr Gly Leu Tyr Ser Ile 375

Tyr Gln Gly Gly Ile Phe Glu Leu Lys Glu Asn Asp Arg Ile Phe Val

Ser Val Thr Asn Glu His Leu Ile Asp Met Asp His Glu Ala Ser Phe 410

Phe Gly Ala Phe Leu Val Gly 420

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gac Asp	atc Ile 15	att Ile	gaa Glu	tgg Trp	aca Thr	acc Thr 20	cag Gln	aag Lys	ctt Leu	gag Glu	ecc Pro 25	aaa Lys	tct Ser	tgt Cys	gac Asp	97
aaa Lys 30	act Thr	cac His	aca Thr	tgc Cys	cca Pro 35	ccg Pro	tgc Cys	cca Pro	gca Ala	cct Pro 40	gaa Glu	ctc Leu	ctg Leu	Gly ggg	gga Gly 45	145
ccg Pro	tca Ser	gtc Val	ttc Phe	ctc Leu 50	ttc Phe	ecc Pro	cca Pro	aaa Lys	ccc Pro 55	aag Lys	gac Asp	acc Thr	ctc Leu	atg Met 60	atc Ile	193
tcc Ser	cgg Arg	acc Thr	cct Pro 65	gag Glu	gtc Val	aca Thr	tgc Cys	gtg Val 70	gtg Val	gtg Val	gac Asp	gtg Val	agc Ser 75	cac His	gaa Glu	241
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aat Asn	gcc Ala 95	aag Lys	aca Thr	aag Lys	ccg Pro	cgg Arg 100	gag Glu	gag Glu	cag Gln	tac Tyr	aac Asn 105	agc Ser	acg Thr	tac Tyr	cgt Arg	337
gtg Val 110	gtc Val	agc Ser	gtc Val	ctc Leu	acc Thr 115	gtc Val	ctg Leu	cac His	cag gac tgg ctg aat ggc aag Gln Asp Trp Leu Asn Gly Lys 120  gcc ctc cca gcc ccc atc gag Ala Leu Pro Ala Pro Ile Glu 135  136		385					
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gag Glu 190	Ser	aat Asn	Gly ggg	cag Gln	ccg Pro 195	gag Glu	aac Asn	aac Asn	tac Tyr	aag Lys 200	acc Thr	acg Thr	cct Pro	ccc Pro	gtg Val 205	625
ctg Leu	gac Asp	tcc Ser	gac Asp	ggc Gly 210	tcc Ser	ttc Phe	ttc Phe	ctc Leu	tac Tyr 215	agc Ser	aag Lys	ctc Leu	acc Thr	gtg Val 220	gac Asp	673
aag Lys	agc Ser	agg Arg	tgg Trp 225	cag Gln	cag Gln	Gly	aac Asn	gtc Val 230	ttc Phe	tca Ser	tgc Cys	tcc Ser	gtg Val 235	atg Met	cat His	721
gag	gct	ctg	cac	aac	cac	tac	acg	cag	aag	agc	ctc	tcc	ctg	tct	ccg	769

Glu	Ala	Leu 240	His	Asn	His	Tyr	Thr 245	Gln	Lys	Ser	Leu	Ser 250	Leu	Ser	Pro	
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	act Thr															865
	att Ile						_	_			-					913
	aag Lys	_		_					_		_					961
	cgg Arg															1009
	ctg Leu 335															1057
	ttc Phe															1105
	aag Lys		-			_	-	_								1153
	tat Tyr									-						1201
	tcc Ser	-		_				-								1249
	ttc Phe 415															1297
	cat His															1345
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325 330 335

Ile Glu Glu Gly Leu Tyr Tyr Ile Tyr Ser Gln Thr Tyr Phe Arg
340 345 350

Phe Gln Glu Ala Glu Asp Ala Ser Lys Met Val Ser Lys Asp Lys Val 355 360 365

Arg Thr Lys Gln Leu Val Gln Tyr Ile Tyr Lys Tyr Thr Ser Tyr Pro 370 380

Asp Pro Ile Val Leu Met Lys Ser Ala Arg Asn Ser Cys Trp Ser Arg 385 390 395 400

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405 410 415

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Asp Ile Ile Glu Trp Thr Thr Gln Lys Leu Glu Pro Lys Ser Cys Asp
15 20 25

aaa act cac aca tgc cca ccg tgc cca gca cct gaa ctc ctg ggg gga 145 Lys Thr His Thr Cys Pro Pro Cys Pro Ala Pro Glu Leu Leu Gly Gly 30 45

ccg tca gtc ttc ctc ttc ccc cca aaa ccc aag gac acc ctc atg atc 193
Pro Ser Val Phe Leu Phe Pro Pro Lys Pro Lys Asp Thr Leu Met Ile
50 55 60

tee egg ace eet gag gte aca tge gtg gtg gtg gae gtg age eae gaa 241 Ser Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Val Ser His Glu

gac cct gag gtc aag ttc aac tgg tac gtg gac ggc gtg gag gtg cat 289
Asp Pro Glu Val Lys Phe Asn Trp Tyr Val Asp Gly Val Glu Val His
80 85 90

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														atc Ile 140		433
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_												ag ctc acc gtg gac Lys Leu Thr Val Asp 220  egc tcc gtg atg cat Lys Ser Val Met His 235				721
														tct Ser		769
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		-	_			_						_	_	gga Gly		865
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														atc Ile		961
														ttt Phe		1009
gaa	gct	gaa	gac	gct	tcc	aag	atg	gtc	tca	aag	gac	aag	gtg	aga	acc	1057

Glu Ala Glu Asp Ala Ser Lys Met Val Ser Lys Asp Lys Val Arg Thr 335 340 345	
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ata gtg ctc atg aag agc gcc aga aac agc tgt tgg tcc aga gat gcc Ile Val Leu Met Lys Ser Ala Arg Asn Ser Cys Trp Ser Arg Asp Ala 370 375 380	1153
gag tac gga ctg tac tcc atc tat cag gga gga ttg ttc gag cta aaa Glu Tyr Gly Leu Tyr Ser Ile Tyr Gln Gly Gly Leu Phe Glu Leu Lys 385 390 395	
aaa aat gac agg att ttt gtt tct gtg aca aat gaa cat ttg atg gac Lys Asn Asp Arg Ile Phe Val Ser Val Thr Asn Glu His Leu Met Asp 400 405 410	1249
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- 23 -

Pro	Ser	Arg	Asp	Glu 165	Leu	Thr	Lys	Àsn	Gln 170	Val	Ser	Leu	Thr	Cys 175	Leu																																																									-		
Val	Lys	Gly	Phe 180	Tyr	Pro	Ser	Asp	Ile 185	Ala	Val	Glu	Trp	Glu 190	Ser	Asn																																																											
Gly	Gln	Pro 195	Glu	Asn	Asn	Tyr	Lys 200	Thr	Thr	Pro	Pro	Val 205	Leu	Asp	Ser																																																											
Asp	Gly 210	Ser	Phe	Phe	Leu	Tyr 215	Ser	Lys	Leu	Thr	Val 220	Asp	Lys	Ser	Arg																																																											
Trp 225	Gln	Gln	Gly	Asn	Val 230	Phe	Ser	Cys	Ser	Val 235	Met	His	Glu	Ala	Leu 240																																																											
His	Asn	His	Tyr	Thr 245	Gln	Lys	Ser	Leu	Ser 250	Leu	Ser	Pro	Gly	Lys 255	Gly																																																											
Gly	Arg	Pro	Gln 260	Lys	Val	Ala	Ala	His 265	Ile	Thr	Gly	Ile	Thr 270	Arg	Arg																																																											
Ser	Asn	Ser 275	Ala	Leu	Ile	Pro	Ile 280	Ser	Lys	Asp	Gly	Lys 285	Thr	Leu	Gly																																																											
Gln	Lys 290	Ile	Glu	Ser	Trp	Glu 295	Ser	Ser	Arg	Lys	Gly 300	His	Ser	Phe	Leu																																																											
Asn 305	Hi.s	Val	Leu	Phe	Arg 310	Asn	Gly	Glu Leu Val Ile Glu Gln Glu Gly 315 320																																																																		
Leu	Tyr	Tyr	Ile	Tyr 325	Ser	Gln	Thr	Tyr	Phe 330	Arg	Phe	Gln	Glu	Ala 335	Glu																																																											
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Leu 385	Tyr	Ser	Ile	Туг	Gln 390	Gly	Gly	Leu	Phe	Glu 395	Leu	Lys	Lys	Asn	Asp 400																																																											
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Glu	Ala	Ser	Phe 420	Phe	Gly	Ala	Phe	Leu 425	Ile																																																																	

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## INTERNATIONAL SEARCH REPORT

ir ational Application No PCT/US 00/08004

A. CLASSIFICATION OF SUBJECT MATTER
IPC 7 CO7K19/00 CO7K14/705 C12N15/86 C12N5/10 A61K38/18 A61P37/00 A61P31/12 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) IPC 7 CO7K Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) STRAND, MEDLINE, CANCERLIT, AIDSLINE, LIFESCIENCES, CHEM ABS Data, WPI Data, EPO-Internal, PAJ C. DOCUMENTS CONSIDERED TO BE RELEVANT Category 5 Citation of document, with indication, where appropriate, of the relevant passages Relevant to claim No. X WO 97 01633 A (IMMUNEX CORP) 1 - 1916 January 1997 (1997-01-16) page 9, line 11 -page 13, line 8 page 21, line 5 -page 24, line 2 examples 8,9,11 claims X WO 97 46686 A (AMGEN INC) 1,3,6,7, 11 December 1997 (1997-12-11) 9,12-17, cited in the application page 3, line 13-24 claims 14-19 page 10, line 2-31 -/---ΧI Further documents are listed in the continuation of box C. Patent family members are listed in annex. \* Special categories of cited documents : "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "E" earlier document but published on or after the International "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone filing date "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such docu-\*O\* document referring to an oral disclosure, use, exhibition or other means ments, such combination being obvious to a person skilled "P" document published prior to the international filing date but later than the priority date claimed "&" document member of the same patent family Date of the actual completion of the international search Date of mailing of the International search report 5 July 2000 27/07/2000 Name and mailing address of the ISA Authorized officer European Patent Office, P.B. 5818 Patentlaan 2 NL -- 2280 HV Rijewijk Tel. (+31--70) 340--2040, Tx. 31 651 epo ni, Fax: (+31--70) 340--3016 Covone, M

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Ir ational Application No PCT/US 00/08004

		FC1/03 00/08004
	lation) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	SHERIDAN J P ET AL: "Control of TRAIL-induced apoptosis by a family of signaling and decoy receptors" SCIENCE, US, AMERICAN ASSOCIATION FOR THE ADVANCEMENT OF SCIENCE, vol. 277, 8 August 1997 (1997-08-08), pages 818-821, XP002065023 ISSN: 0036-8075 the whole document	1-19
A	KIM J -K ET AL: "IDENTIFYING AMINO ACID RESIDUES THAT INFLUENCE PLASMA CLEARANCE OF MURINE IGG1 FRAGMENTS BY SITE-DIRECTED MUTAGENESIS" EUROPEAN JOURNAL OF IMMUNOLOGY, DE, WEINHEIM, vol. 24, no. 3, 1 January 1994 (1994-01-01), pages 542-548, XP000590871 ISSN: 0014-2980 the whole document	2,8
<b>A</b>	DANILENKO D M (REPRINT) ET AL: "AGP - 1, a novel member of the tumor necrosis factor family, induces hepatic necrosis and inflammation in transgenic mice."  FASEB JOURNAL, (28 FEB 1997) VOL. 11, NO. 3, PP. 2951-2951, XP002141678  the whole document	1-19

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WO	9701633	A	16-01-1997	AU AU CA EP JP NO NZ US	5596599 A 708239 B 6340796 A 2225378 A 0835305 A 11508445 T 976045 A 311982 A 5763223 A	13-01-2000 29-07-1999 30-01-1997 16-01-1997 15-04-1998 27-07-1999 02-03-1998 30-08-1999 09-06-1998
WO	9746686	A	11-12-1997	AU CA EP	3381097 A 2256464 A 0918860 A	05-01-1998 11-12-1997 02-06-1999